

# **Fertoprotective Therapeutics: What is on the horizon for patients?**

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**Northwestern**

# Disclosures

No commercial relationships to disclose

Funding sources

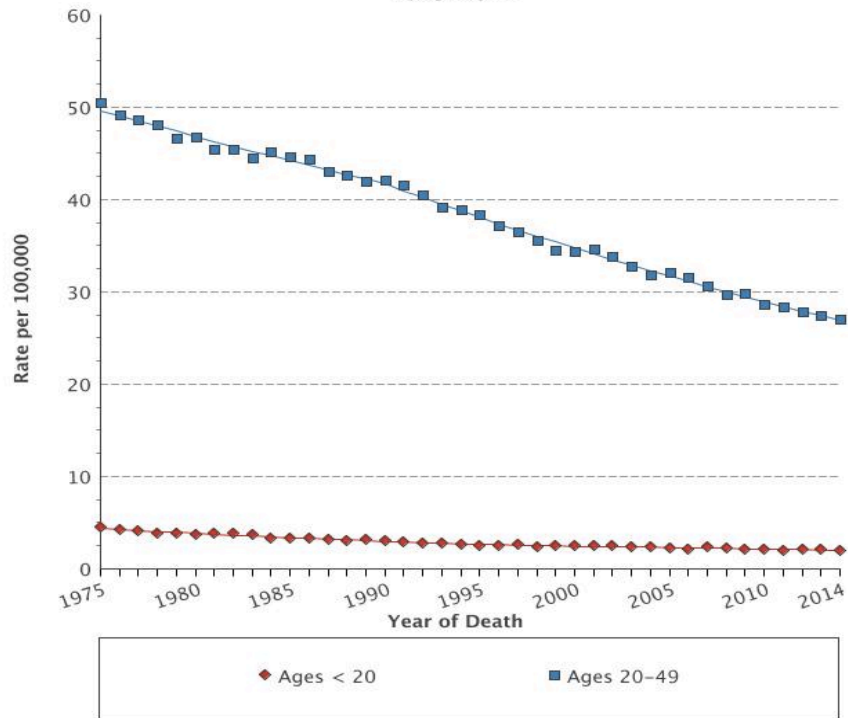
- American Society for Reproductive Medicine
- Foundation for Women's Wellness

# Goals and objectives

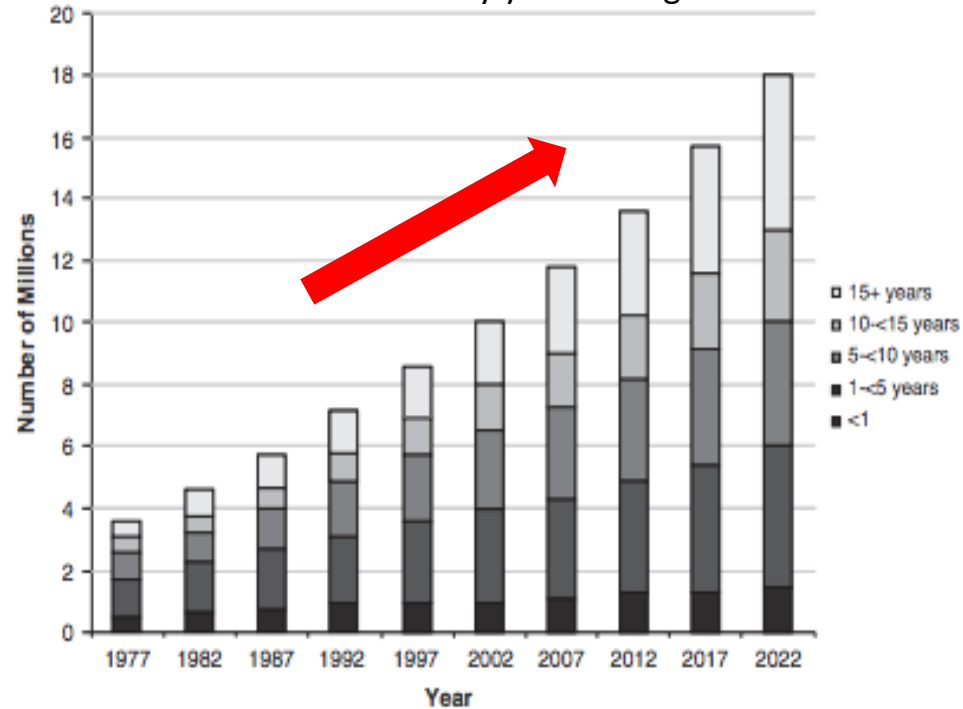
1. Review mechanisms of iatrogenic ovarian injury
2. Discuss limitations of current approaches to fertility preservation
3. Examine experimental approaches on the horizon for pharmacologic fertoprotection

# Cancer survivorship among reproductive-aged women

Age-Adjusted U.S. Mortality Rates  
By Age  
All Sites, All Races, Female  
1975-2014

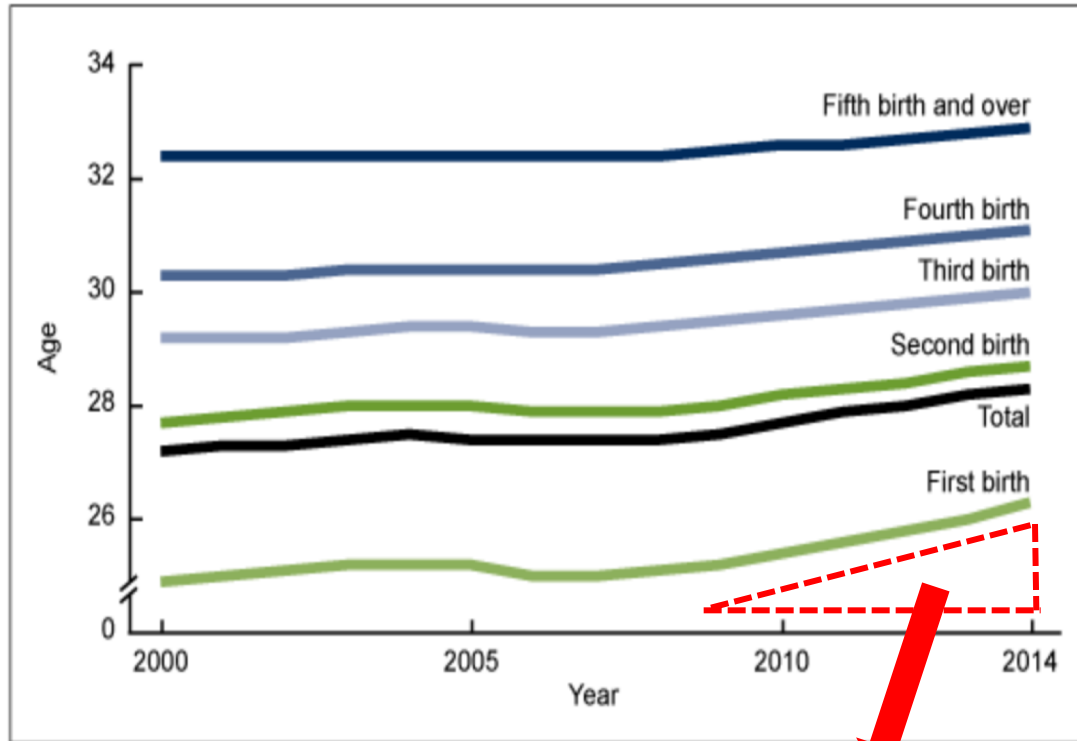


U.S. cancer survivors by year of diagnosis



# Delayed childbearing in the context of a cancer diagnosis

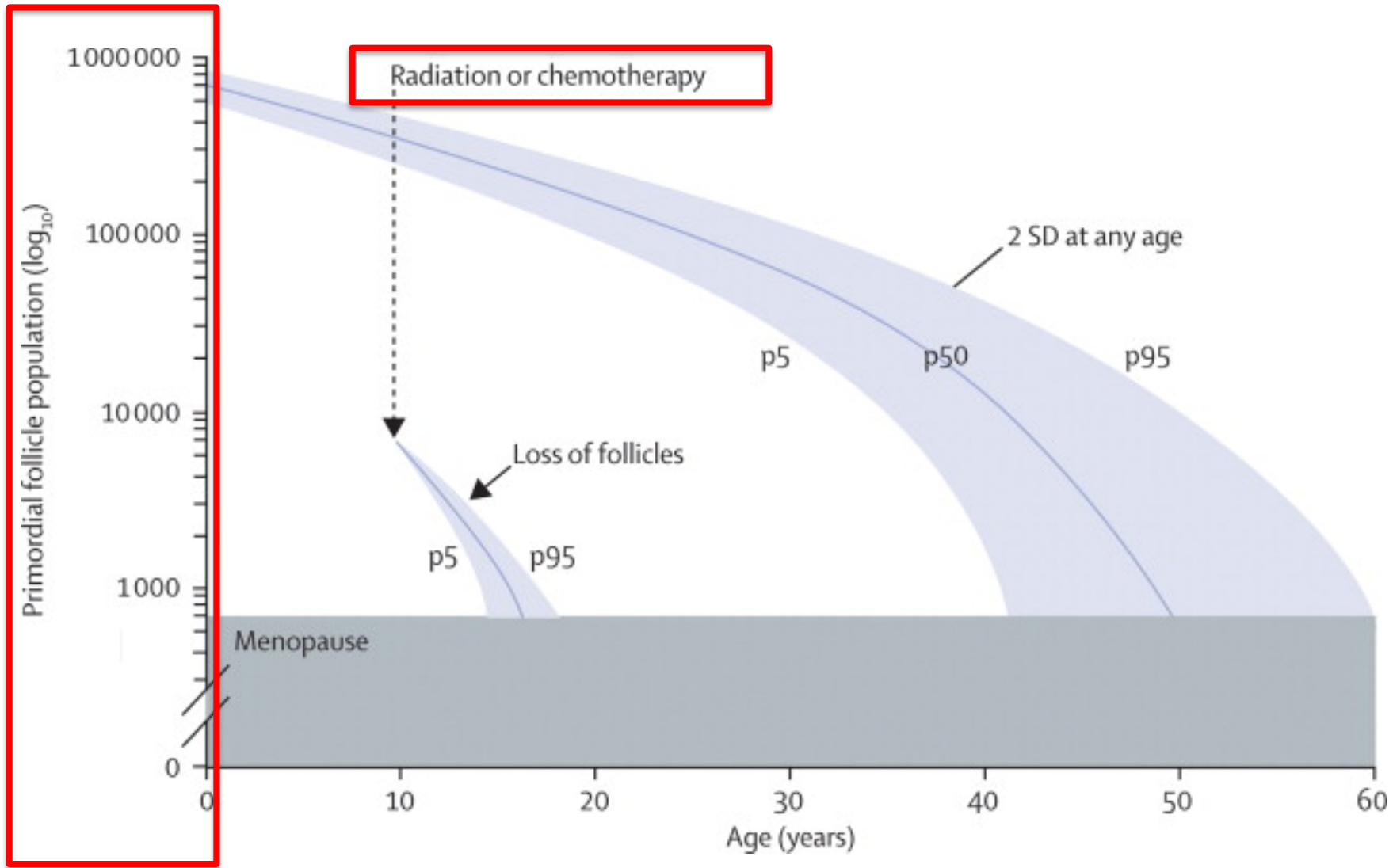
Mean age by birth order: United States, 2000-2014



SOURCE: CDC/NCHS, National Vital Statistics System.

Increasing numbers of women childless at time of cancer diagnosis

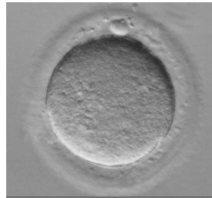
# Impact of treatment on ovarian reserve



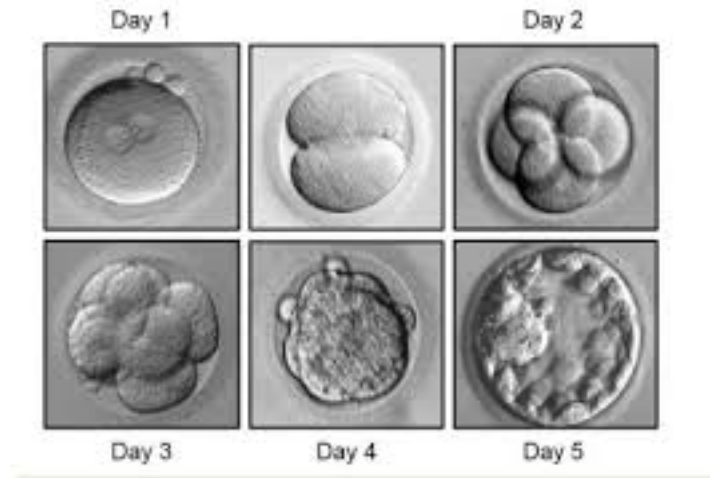
# Options for female fertility preservation

## Established

Oocyte cryopreservation



Embryo cryopreservation



## Investigational/ experimental

Ovarian tissue cryopreservation

Pharmacologic fertoprotection

# Limitations of oocyte/embryo cryopreservation

Hypothetical 35yo woman

Oocyte cryopreservation

Embryo cryopreservation

80% oocyte survival

10 MII oocytes

80% fertilization

8 MII undergo ICSI

30% blastocyst formation

6 2PN



10 MII oocytes

80% fertilization

10 MII oocytes

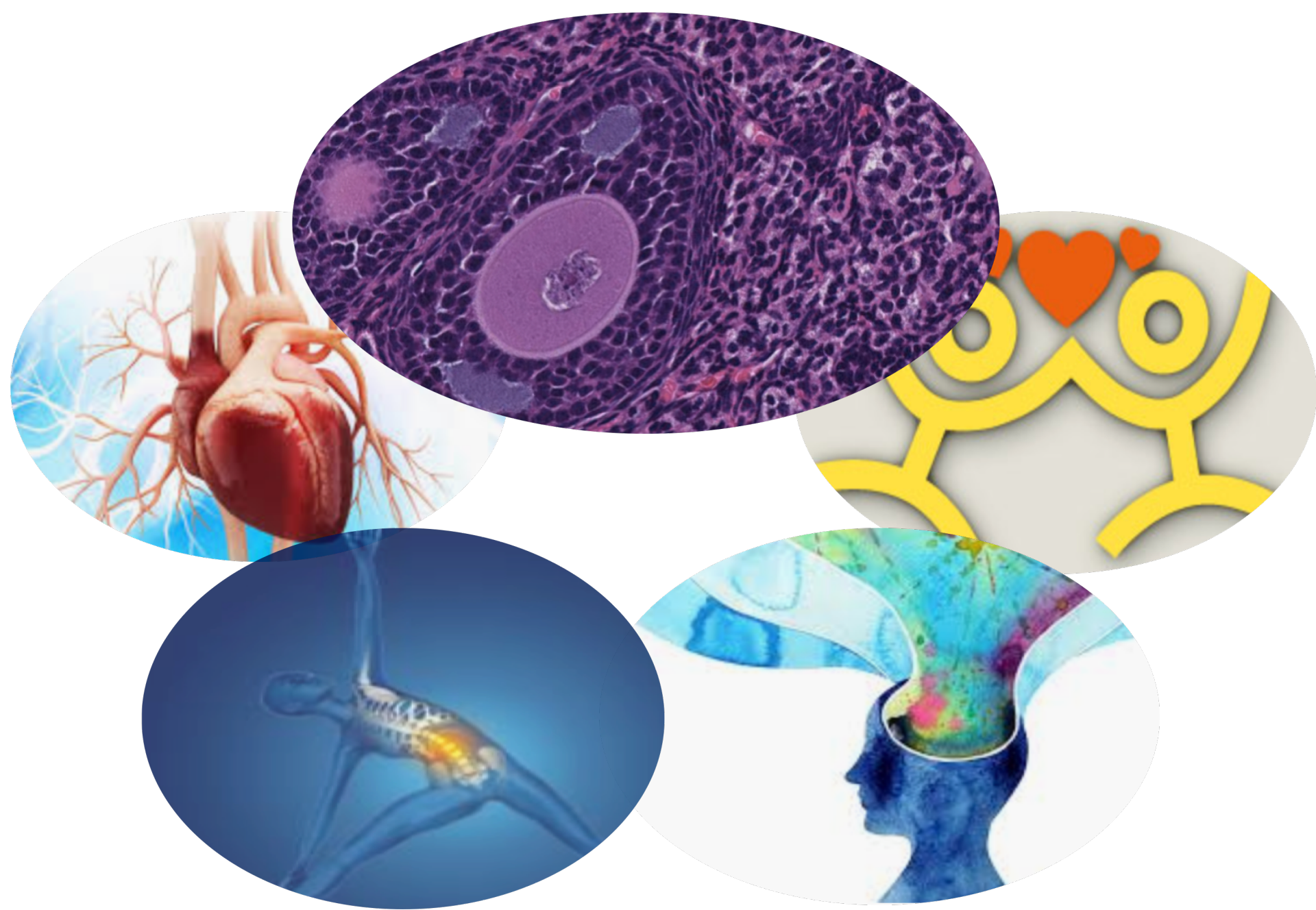
50% blastocyst formation

8 2PN

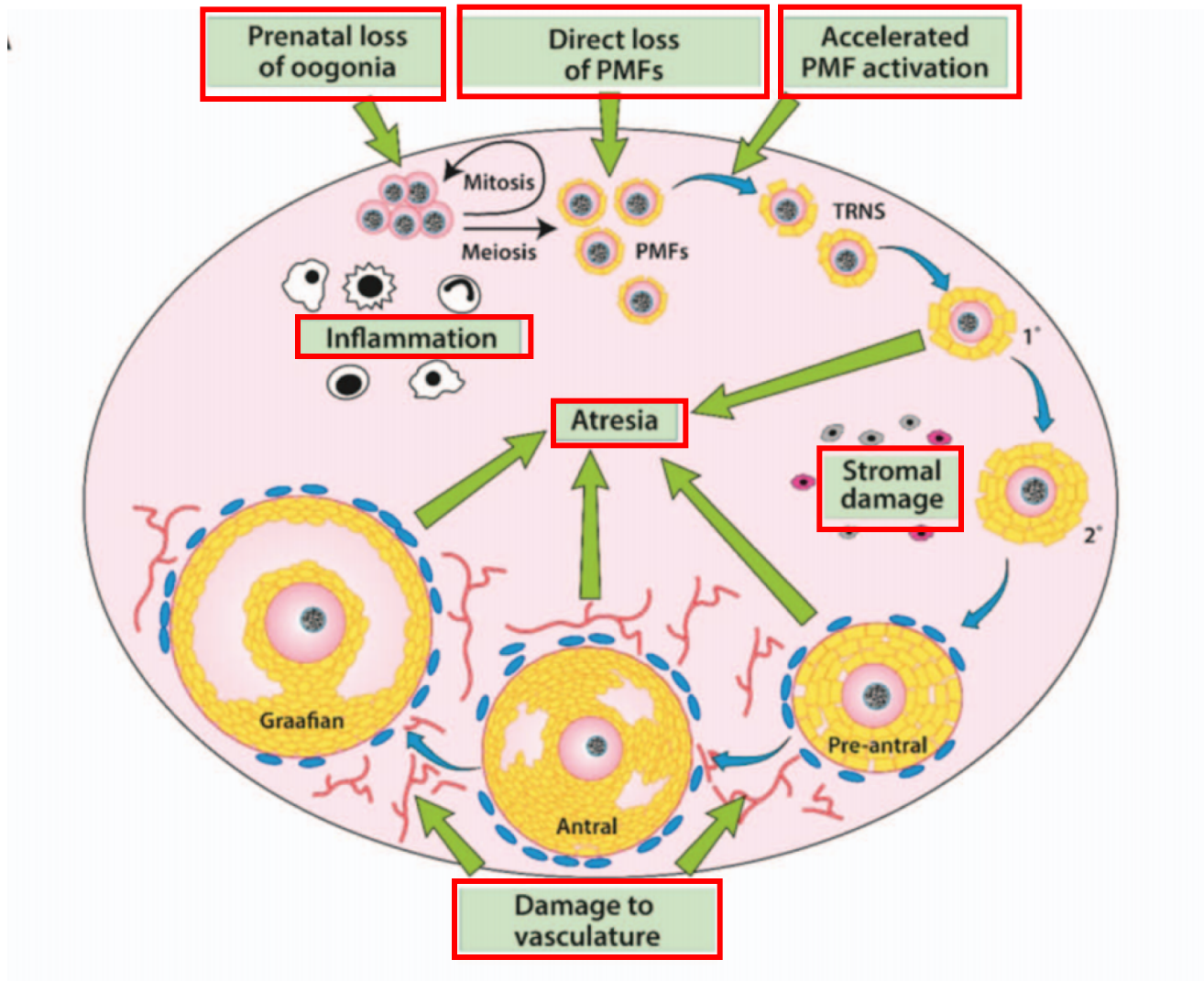


Euploidy?





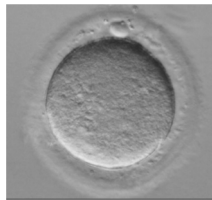
# Mechanisms of iatrogenic ovarian injury



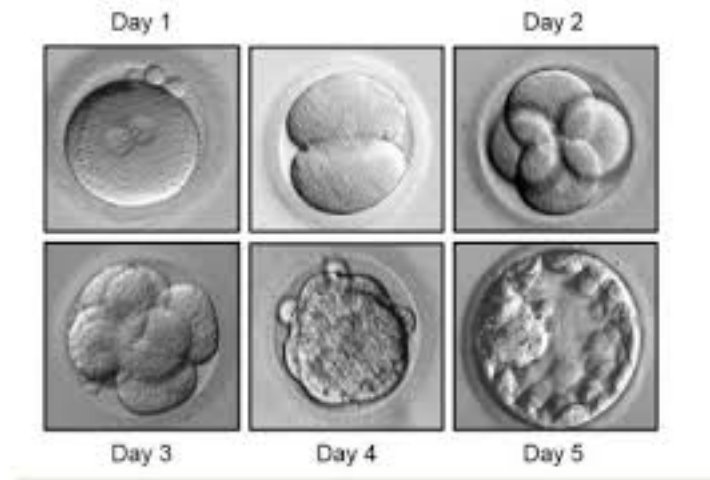
# Options for female fertility preservation

## Established

Oocyte cryopreservation



Embryo cryopreservation

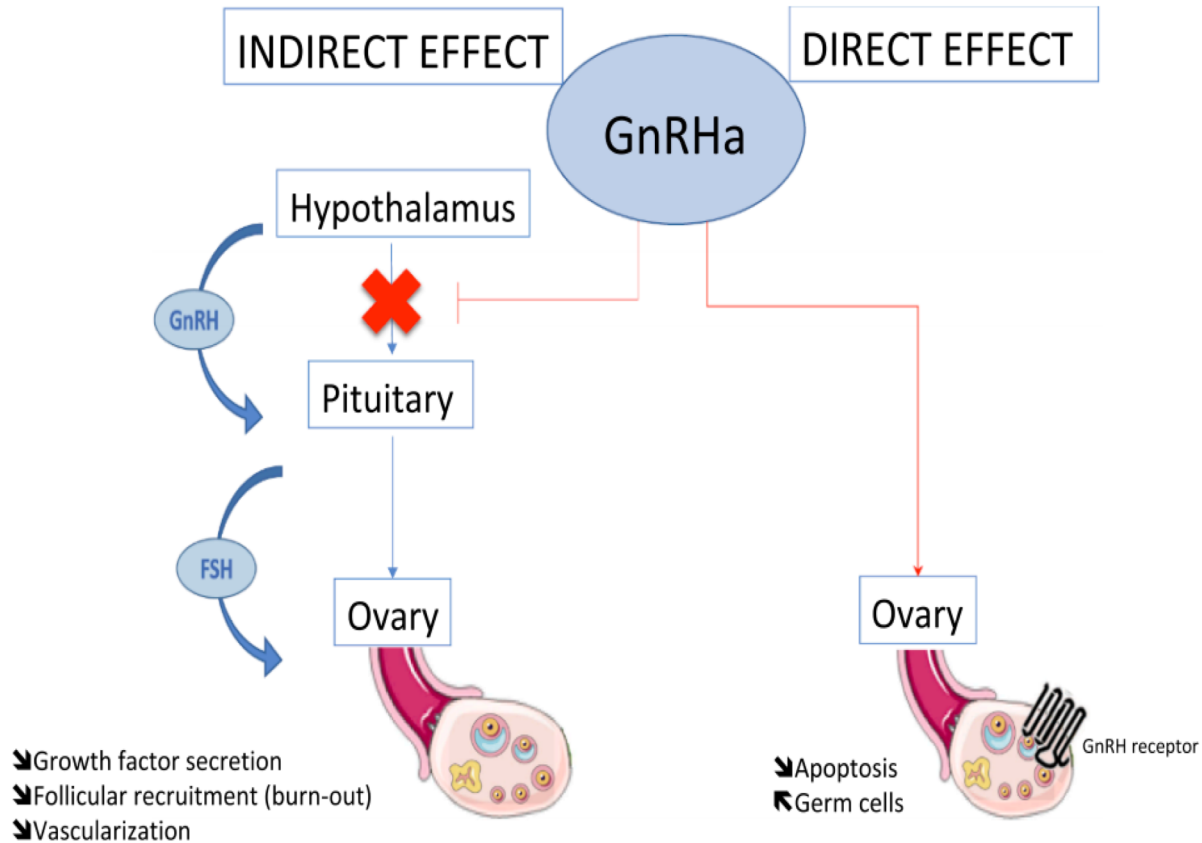


## Investigational/ experimental

Ovarian tissue cryopreservation

Pharmacologic fertoprotection

# GnRH agonist mechanism of action in fertility preservation



# GnRH agonist pre-clinical data: murine models

**Table 1**  
Preclinical studies in female mice evaluating temporary ovarian suppression with GnRH $\alpha$  during chemotherapy.

Authors	Type of gonadotoxic treatment	Main results	Overall results
Yuce et al., 2004	Cyclophosphamide	* Small protection of primordial follicles	Protection (only against high dose of cyclophosphamide)
Danforth et al., 2005; Kishk et al., 2013; Hasky et al., 2015; Kanter et al., 2016	Cyclophosphamide	* Dose-dependant protection of the ovarian reserve * Slight protection of growing follicles * Preservation of AMH levels * Preservation of fertilization rate, early embryo development and improvement of oocyte quality	Protection
Tan et al., 2010	Busulfan	* Protection of primordial and primary follicles	Protection
Lin et al., 2012; Zhang et al., 2013	Cisplatin	* Protection of quiescent and growing follicles * Preservation of AMH levels	Protection
Deti et al., 2014; Horicks et al., 2015; Horicks et al., 2018	Cyclophosphamide	* No difference in proliferation and apoptosis in the ovaries * No protection of quiescent and growing follicles * No protection of FSH and AMH levels * FSH deficiency does not protect ovarian reserve * <i>In vitro</i> exposure to GnRH $\alpha$ does not preserve follicular survival * No difference in proliferation and apoptosis in the ovaries	No protection
Hasky et al., 2015	Doxorubicin	* Compromise vascular recovery * No preservation of AMH levels	No protection
Park et al., 2017	Docetaxel	* Protection of total follicles * Preservation of proliferation within follicles * Decrease of double-strand DNA breaks	Protection

# GnRH agonist pre-clinical data: rat models

**Table 2**  
Preclinical studies in female rats evaluating temporary ovarian suppression with GnRHa during chemotherapy.

Authors	Type of gonadotoxic treatment	Main results	Overall results
Ataya et al., 1985; Ataya et al., 1988; Bokser et al., 1990; Ataya et al., 1993; Knudtson et al., 2017	Cyclophosphamide	<ul style="list-style-type: none"> <li>* Protection of quiescent and growing follicles</li> <li>* Preservation of LH and E2 levels</li> <li>* Preservation of pregnancy, implantation and live birth rates</li> </ul>	Protection
Montz et al., 1991	Cyclophosphamide	<ul style="list-style-type: none"> <li>* Improvement of fertility only with agonist twice a day</li> </ul>	Partial protection
Letterie et al., 2004; Li et al., 2015; Parlakgumus et al., 2015	Cyclophosphamide	<ul style="list-style-type: none"> <li>* No protection of ovarian reserve and growing follicles</li> <li>* No preservation of fertility</li> <li>* Increase in liver, pulmonary and splenic hemorrhage</li> <li>* No preservation of AMH levels</li> </ul>	No protection
Matsuo et al., 2007; Li et al., 2013	Cisplatin	<ul style="list-style-type: none"> <li>* Protection of ovarian reserve</li> <li>* Preservation of cyclicity</li> </ul>	Protection
Ozcelik et al., 2010	Paclitaxel and/or cisplatin	<ul style="list-style-type: none"> <li>* Protection of ovarian reserve (paclitaxel)</li> <li>* No protection of ovarian reserve (cisplatin)</li> </ul>	Protection only against paclitaxel
Wang et al., 2014	5-fluorouracil	<ul style="list-style-type: none"> <li>* Protection of ovarian reserve</li> <li>* Preservation of AMH and FSH levels</li> <li>* Decrease of apoptotic factors</li> </ul>	Protection

# Pre-clinical data: female primates, human models

**Table 3**  
Preclinical studies in female primates and human models evaluating GnRHa effect during chemotherapy.

Authors	Model	Type of gonadotoxic treatment	Main results	Overall results
Ataya et al., 1995	<i>In vivo</i> study in rhesus monkeys	Cyclophosphamide	<ul style="list-style-type: none"> <li>* Protection of ovarian reserve</li> <li>* Preservation of FSH, E2 and P levels</li> <li>* Interruption of cyclicity</li> </ul>	Protection
Imai et al., 2007	<i>In vitro</i> study on human granulosa cells	Doxorubicin	<ul style="list-style-type: none"> <li>* Direct preservation of E2 levels after FSH stimulation</li> </ul>	Protection
Bildik et al., 2015	<i>In vitro</i> study on human granulosa cells and ovarian tissue fragments	Cyclophosphamide Paclitaxel 5-fluorouracil TAC regimen	<ul style="list-style-type: none"> <li>* No protection of ovarian reserve</li> <li>* No preservation of AMH, E2 and P levels</li> <li>* No upregulation of anti-apoptotic genes</li> <li>* No preservation of the vascular density</li> </ul>	No protection

# 30 + years of clinical data in GnRH agonists:

- 14 randomized studies in breast cancer:
  - Potential prolongation of ovarian function/ possible decreased POI
  - No clear benefit in fertility preservation
- 2 RCTs in lymphoma: No benefit
- 12 meta-analyses: Potential benefit in preventing POI
- Clear benefit: menstrual suppression in women at bleeding risk



# Gonadotropin releasing hormone agonists (GnRHa)

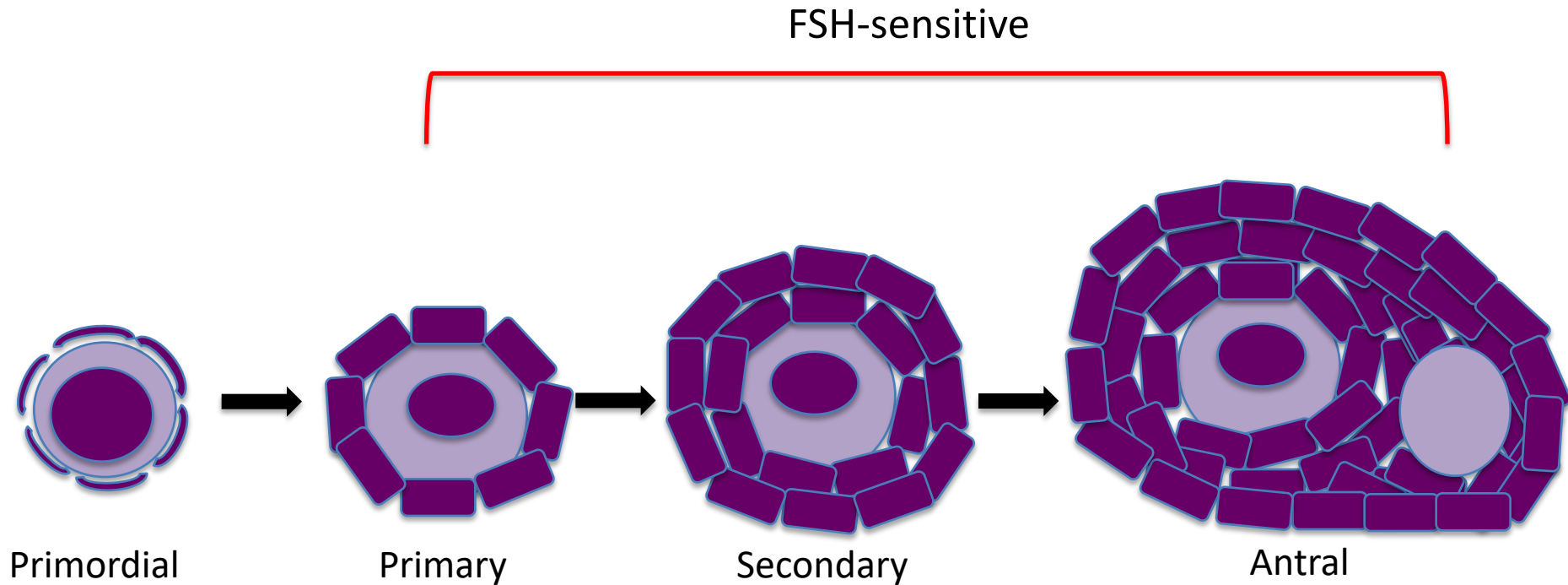
ASCO statement 2013

- Present both embryo and oocyte cryopreservation as established fertility preservation methods
- Discuss the option of ovarian transposition (oophoropexy) when pelvic radiation therapy is performed as cancer treatment
- Inform patients of conservative gynecologic surgery and radiation therapy options
- Inform patients that there is insufficient evidence regarding the effectiveness of ovarian suppression (gonadotropin-releasing hormone analogs) as a fertility preservation method, and these agents should not be relied on to preserve fertility
- Inform patients that other methods (eg, ovarian tissue cryopreservation, which does not require sexual maturity, for the purpose of future transplantation) are still experimental

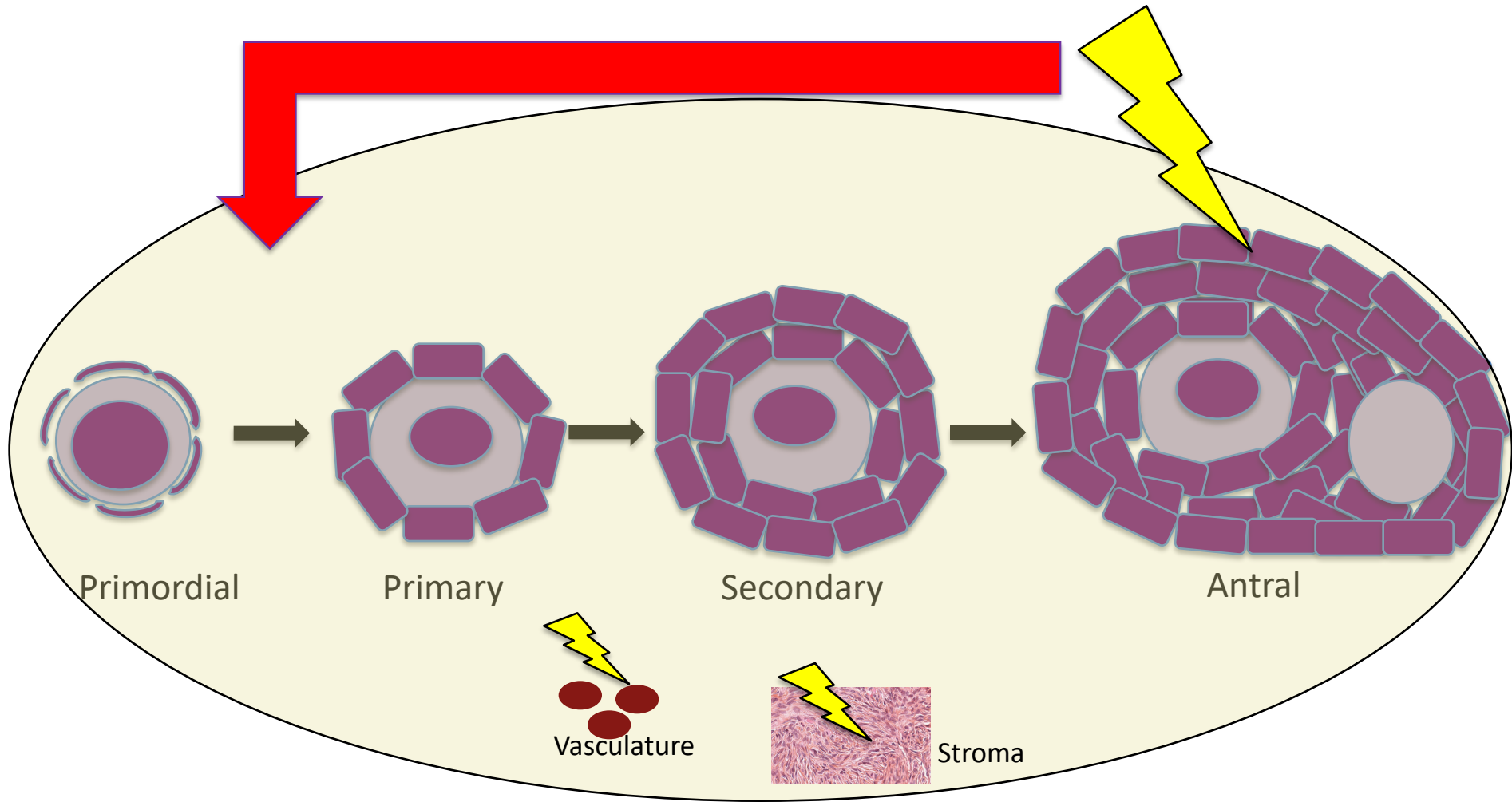
NCCN revised guidelines 2015:

“Randomized trials have shown that suppression with GnRH agonist therapy during adjuvant chemotherapy in **premenopausal women with ER-negative tumors**...may preserve ovarian function and diminish the likelihood of chemotherapy-induced amenorrhea” ... “smaller historical experiences in patients with ER-positive disease... **conflicting results regarding protective effect on fertility**”

# Physiologic ovarian folliculogenesis



# Mechanisms of ovarian damage



# Promoting and inhibitor factors

Promoting

**mTOR**

AKT/PIK3C

Nobox

Kit/KitLG

GDF9

IGF1

GDF9

LH/LHR

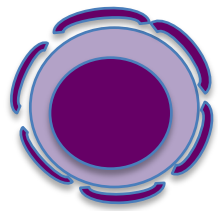
FSH/FSHR

ESR1/2

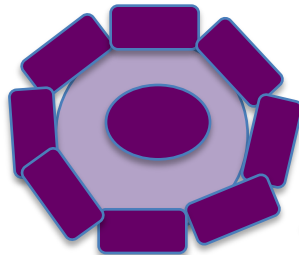
CYP19A1

FMR1

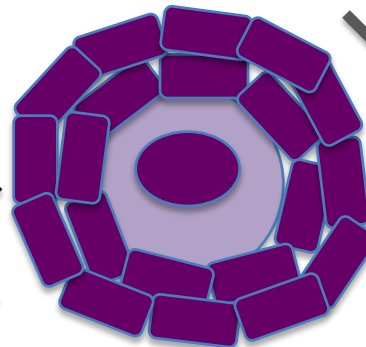
GDF9



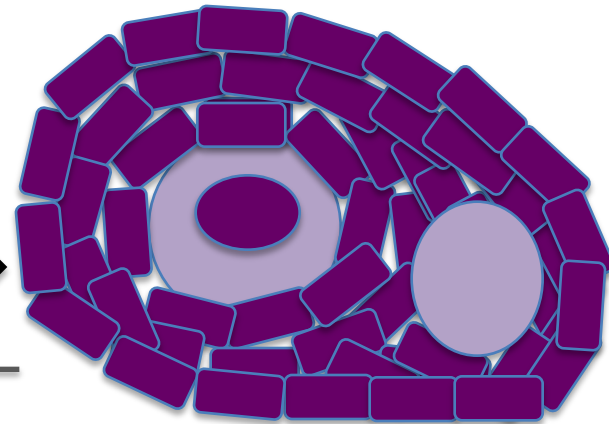
**Primordial**



**Primary**



**Secondary**



**Antral**



Inhibiting

PTEN

S6K1

FOXO3

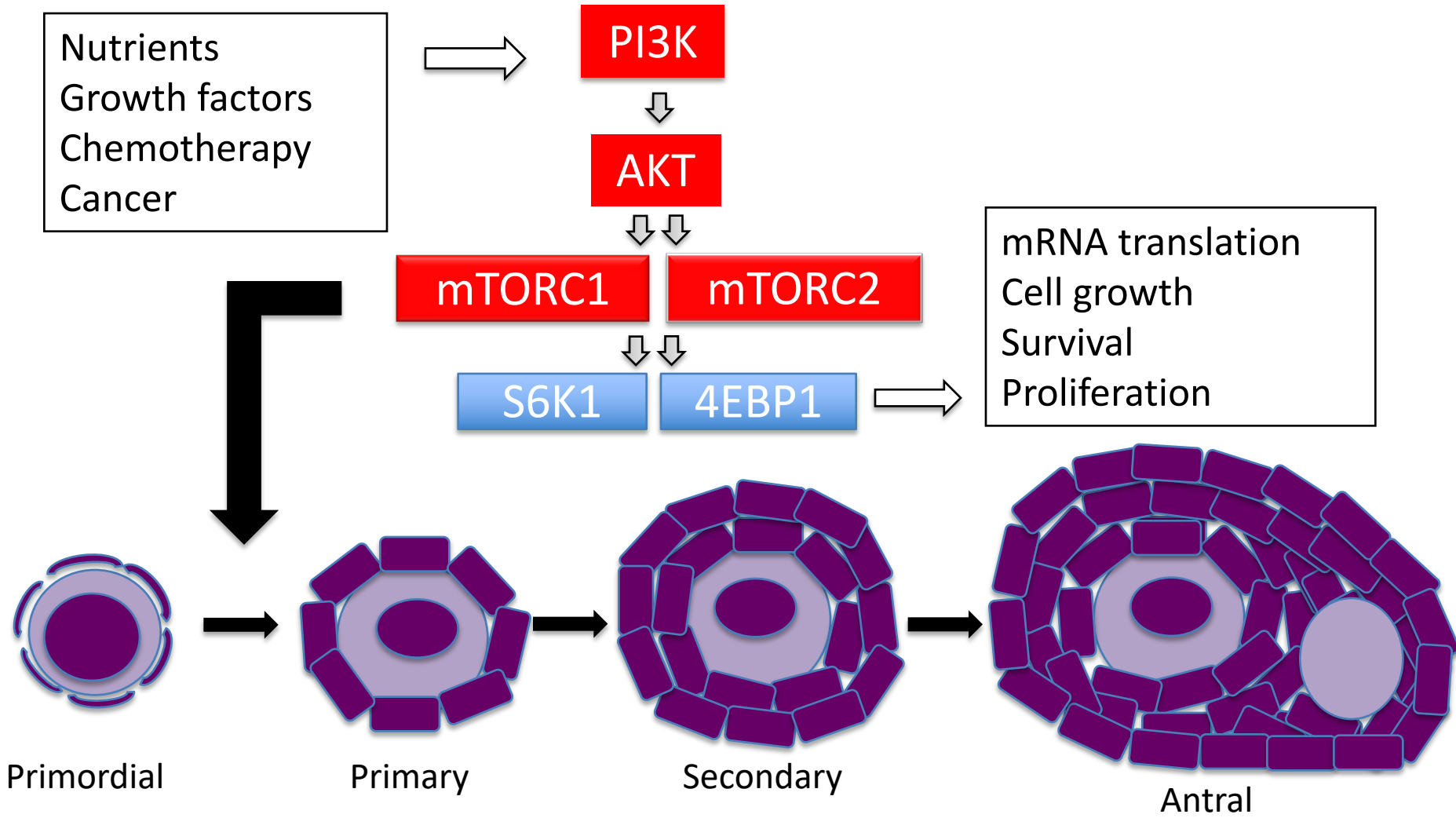
AMH

FOXL2

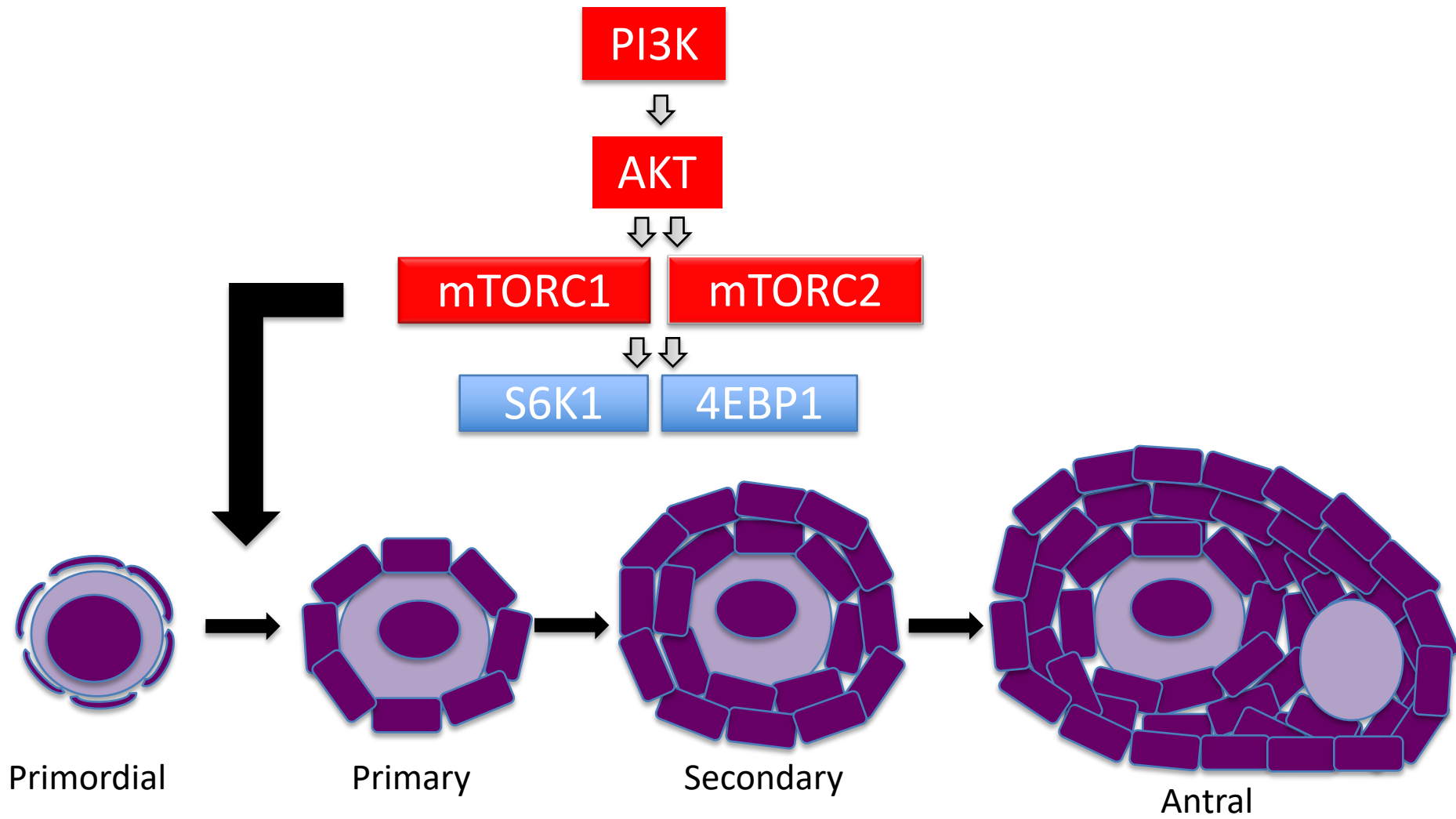
TP53

BAX

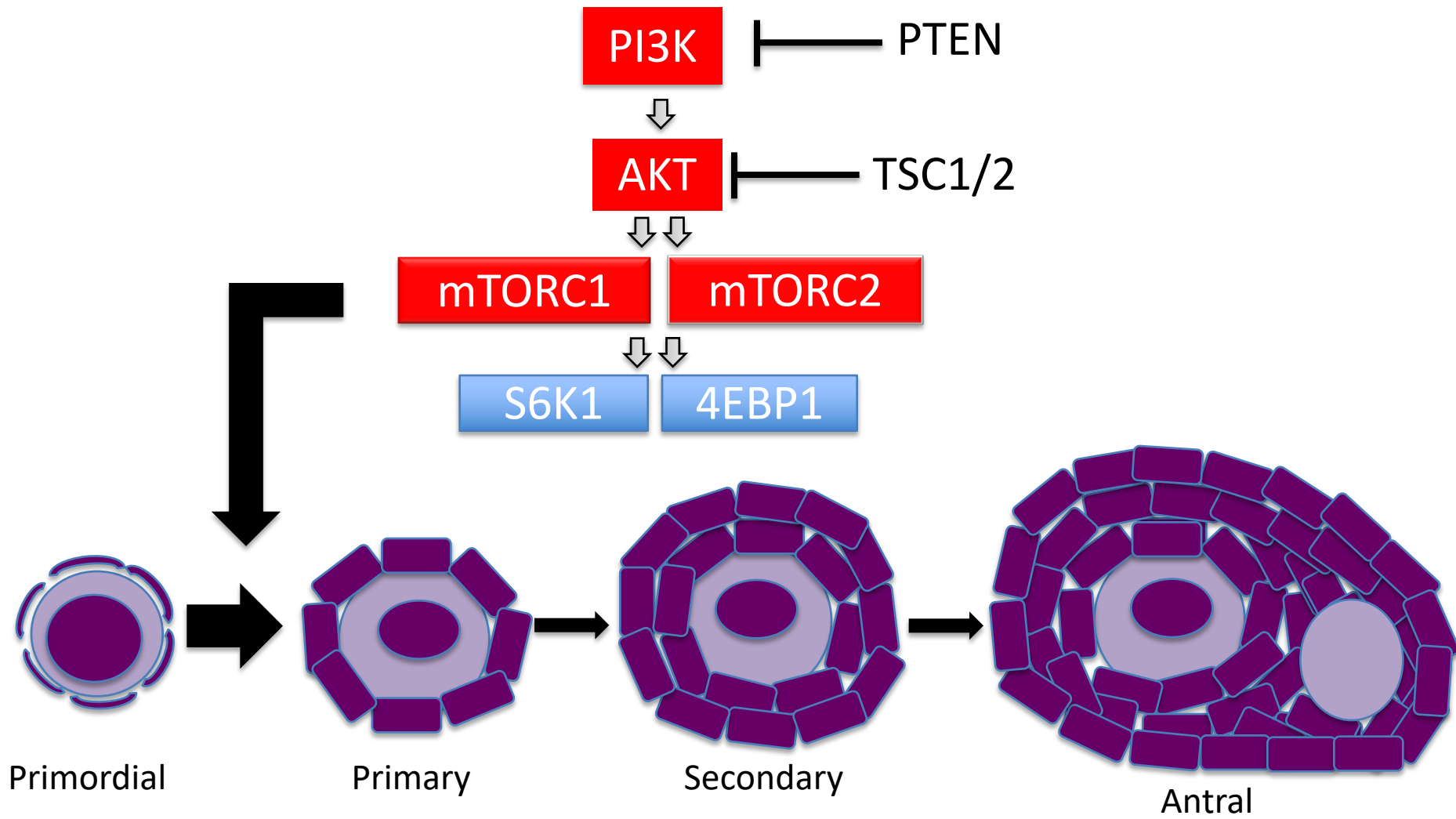
# mTOR pathway



# mTOR pathway critical to primordial follicle activation

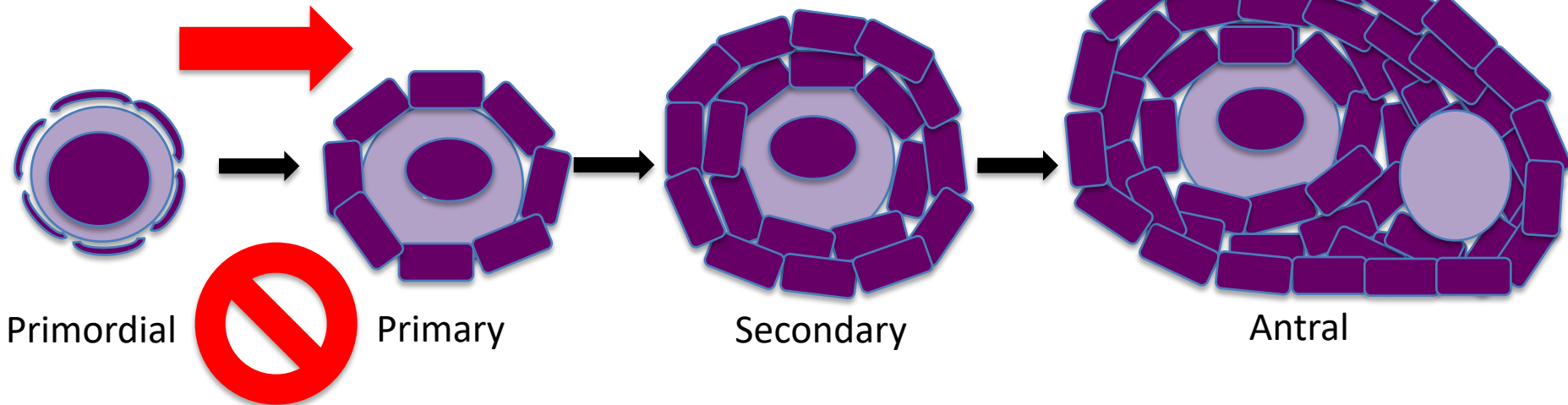


# Up-regulated PI3K/AKT results in follicular depletion



# Potential clinical implications

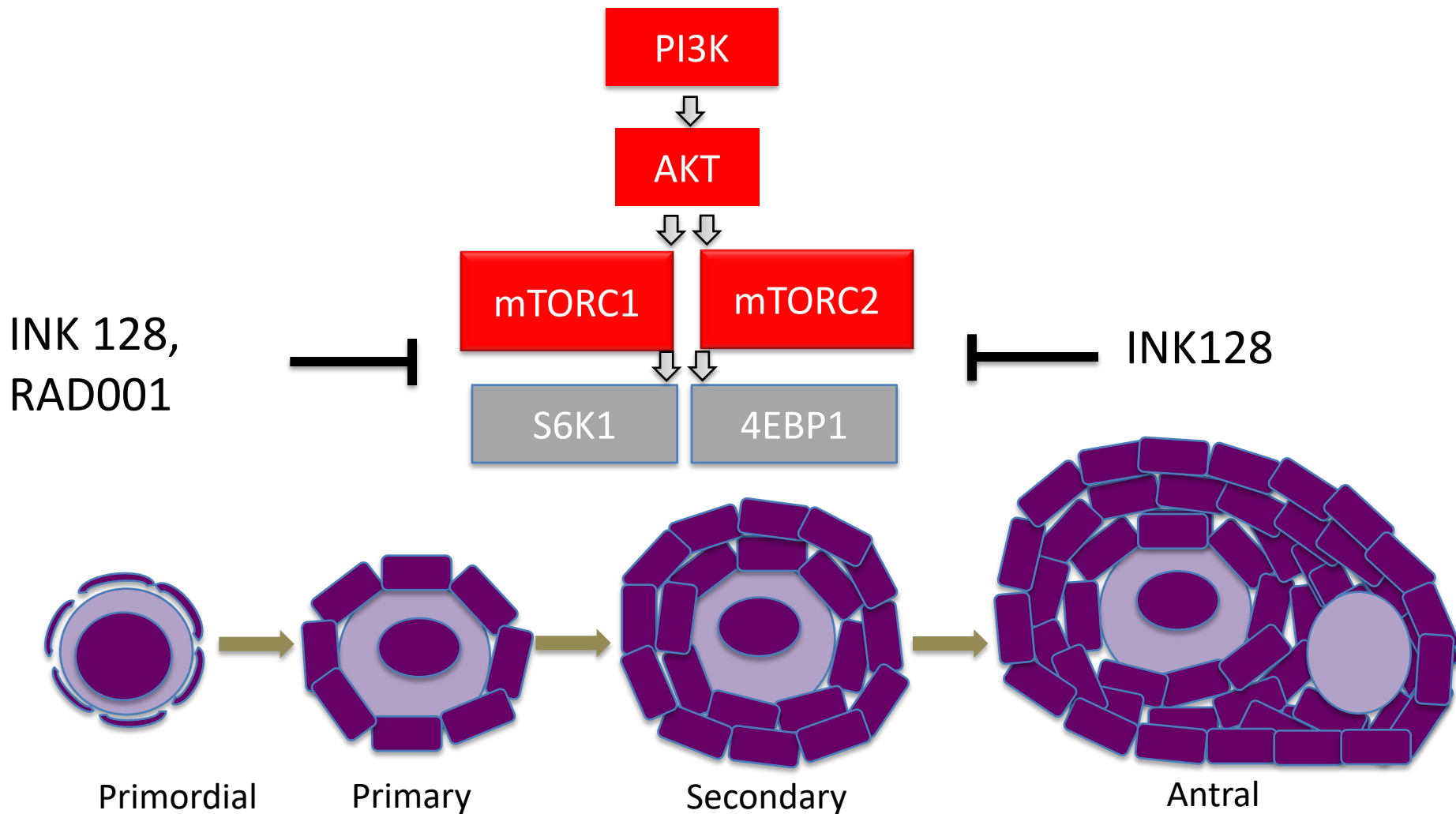
Primary ovarian insufficiency



Fertility preservation  
Ovarian aging



# Hypothesis: mTOR inhibitors preserve ovarian reserve and fertility in mice treated with CY



# mTOR inhibitors widely used for benign and malignant conditions

## mTORC 1 Inhibitors (Everolimus, RAD001)

- **Breast cancer (ER+, HER2 neg) \*\***
- Advanced renal cell carcinoma \*\*
- Subependymal Giant Cell Astrocytoma \*\*
- Tuberous Sclerosis \*\*
- Metastatic pancreatic neuroendocrine tumors \*\*
- Diffuse Large B-Cell Lymphoma
- Epilepsy
- Melanoma
- Cholangiocarcinoma

## mTORC 1/2 (Dual) Inhibitors (INK128, MLN0128)

- Breast cancer
- Neuroblastoma
- Pancreatic cancer
- Renal cell carcinoma
- Thyroid cancer
- Acute lymphoblastic leukemia
- Non-Hodgkin's Lymphoma

\*\*FDA approved: trade name Afinitor (Novartis)



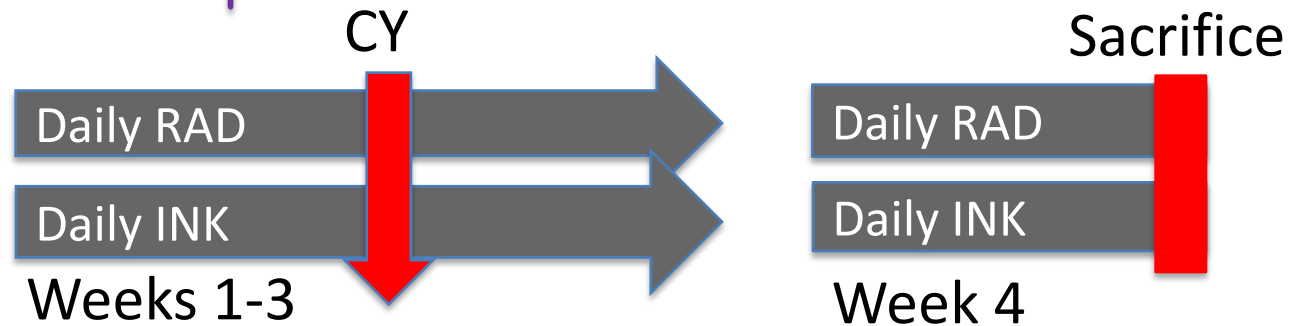
C57BL/6  
8 weeks

Control (PVP) \*  
RAD001 2.5mg/kg \* (mTORC1 inhibitor)  
INK128 0.3 mg/kg \* (mTORC1/2 inhibitor)  
Cyclophosphamide (CY 75mg/kg)  
RAD + CY  
INK + CY

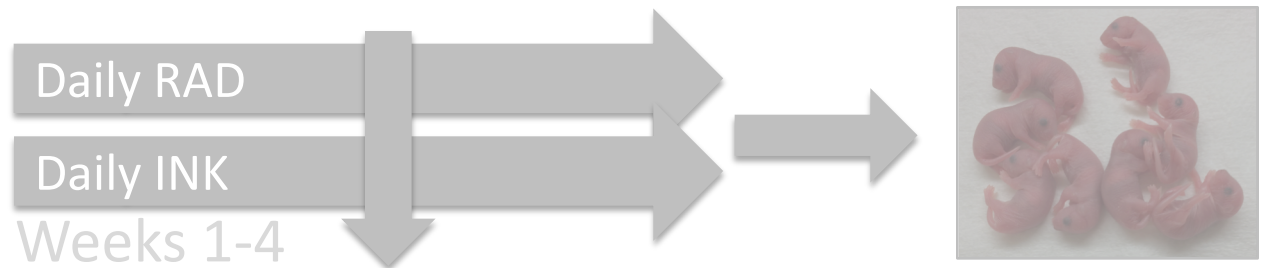


\* Oral gavage

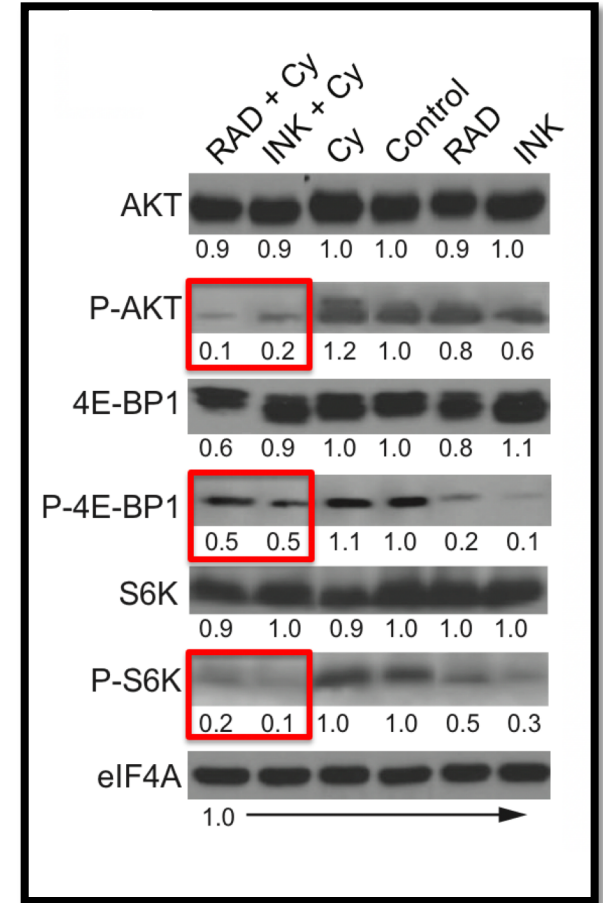
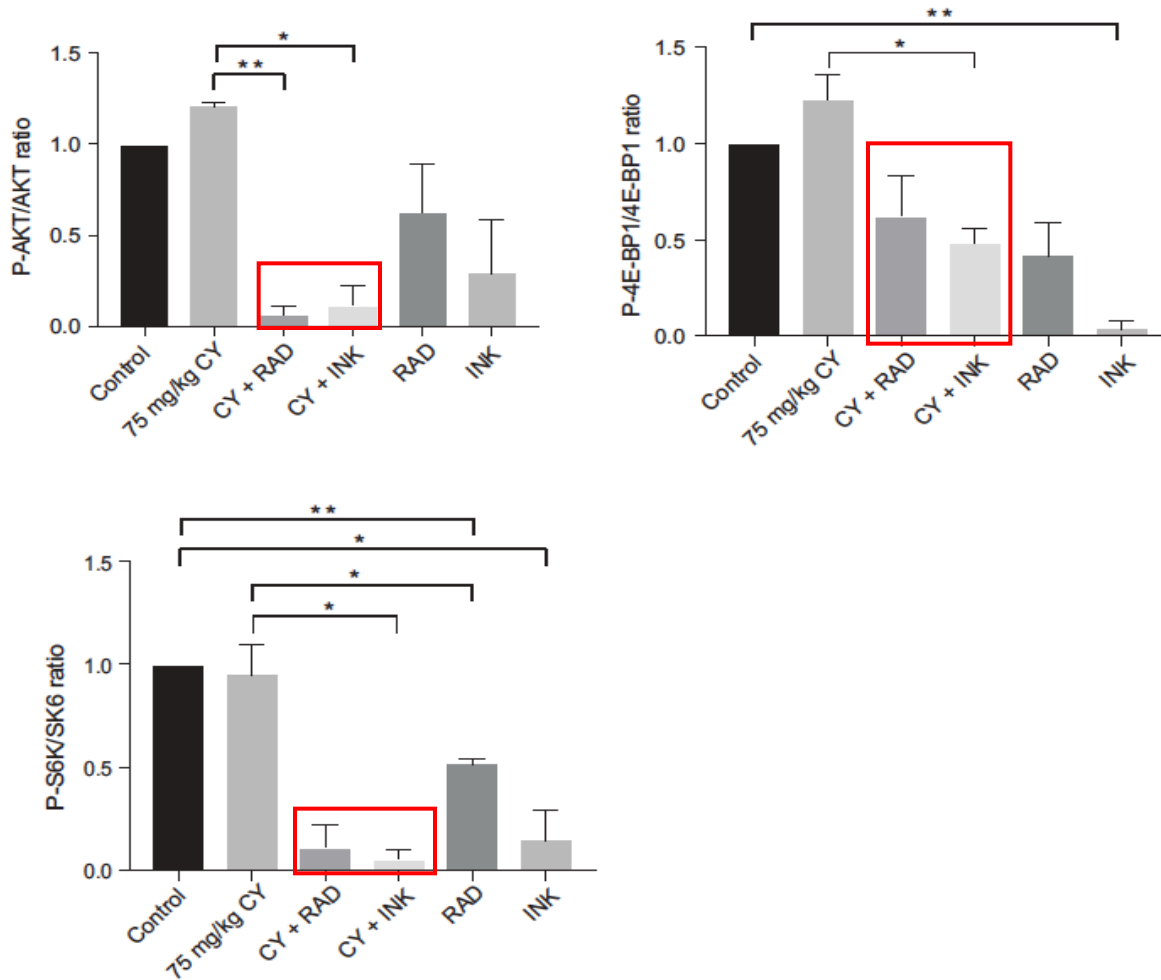
**Aim 1:  
Ovarian reserve**



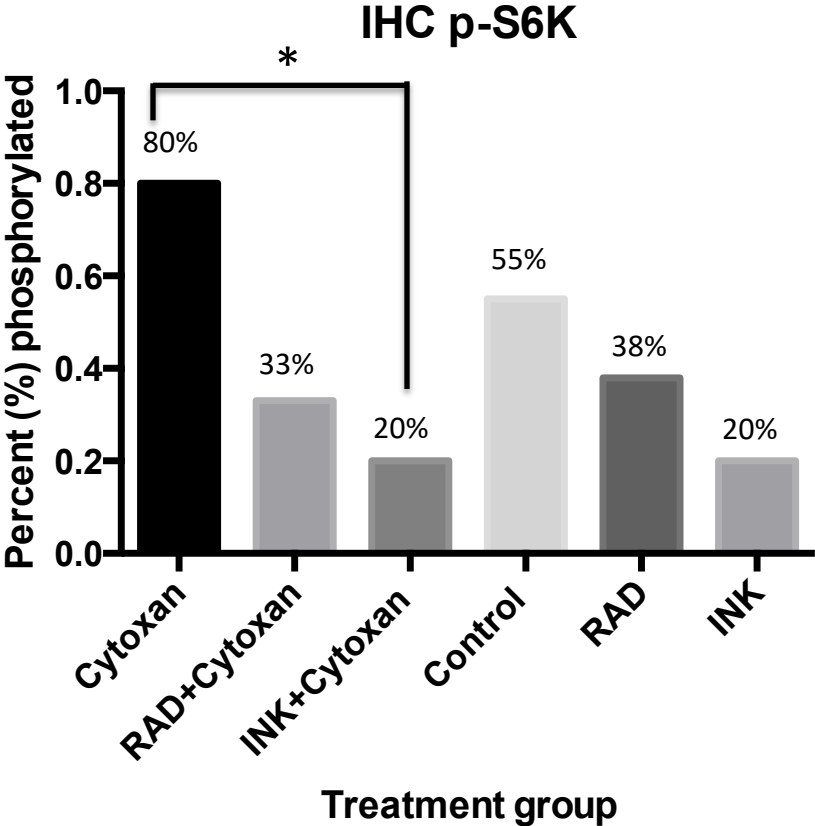
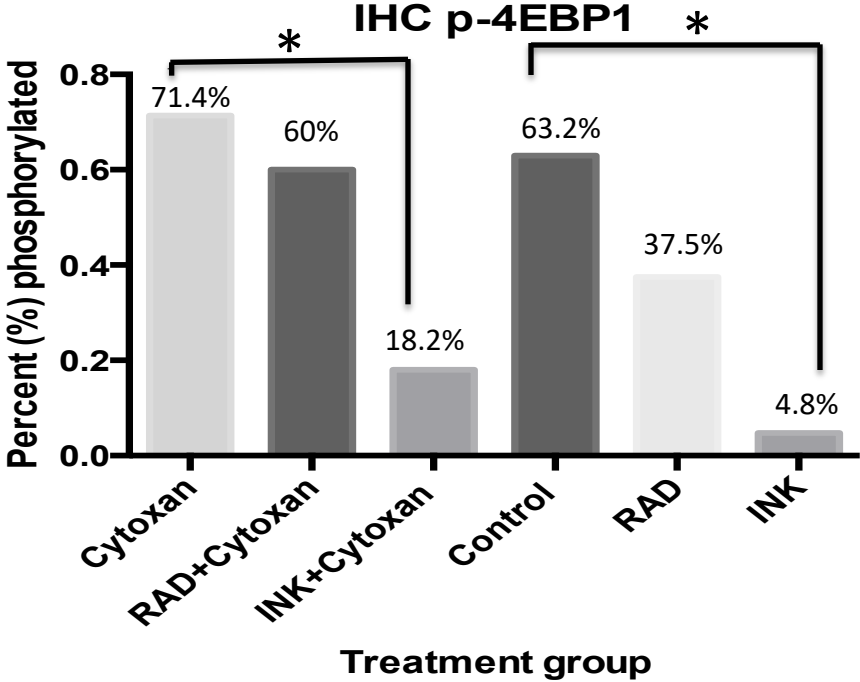
**Aim 2:  
Fertility**



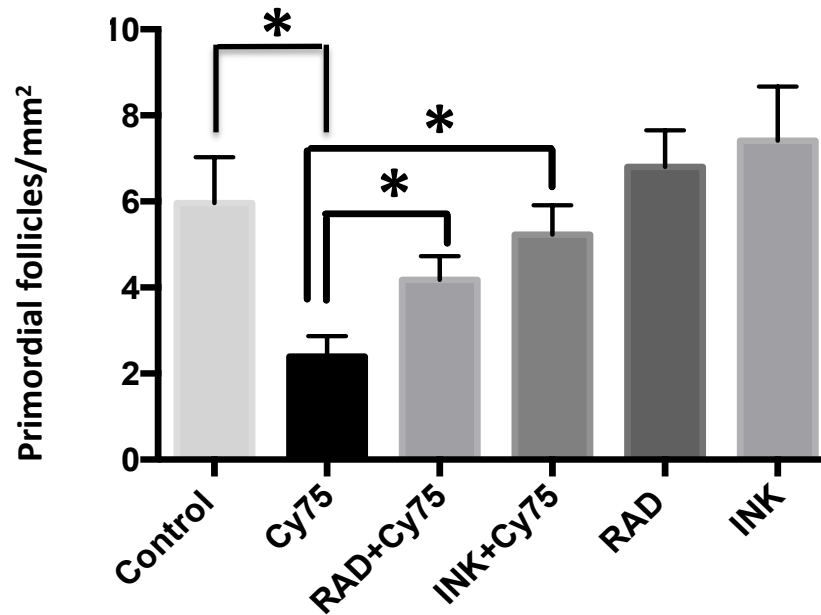
# Down-regulation of mTOR activity in whole ovary lysates of co-treated mice



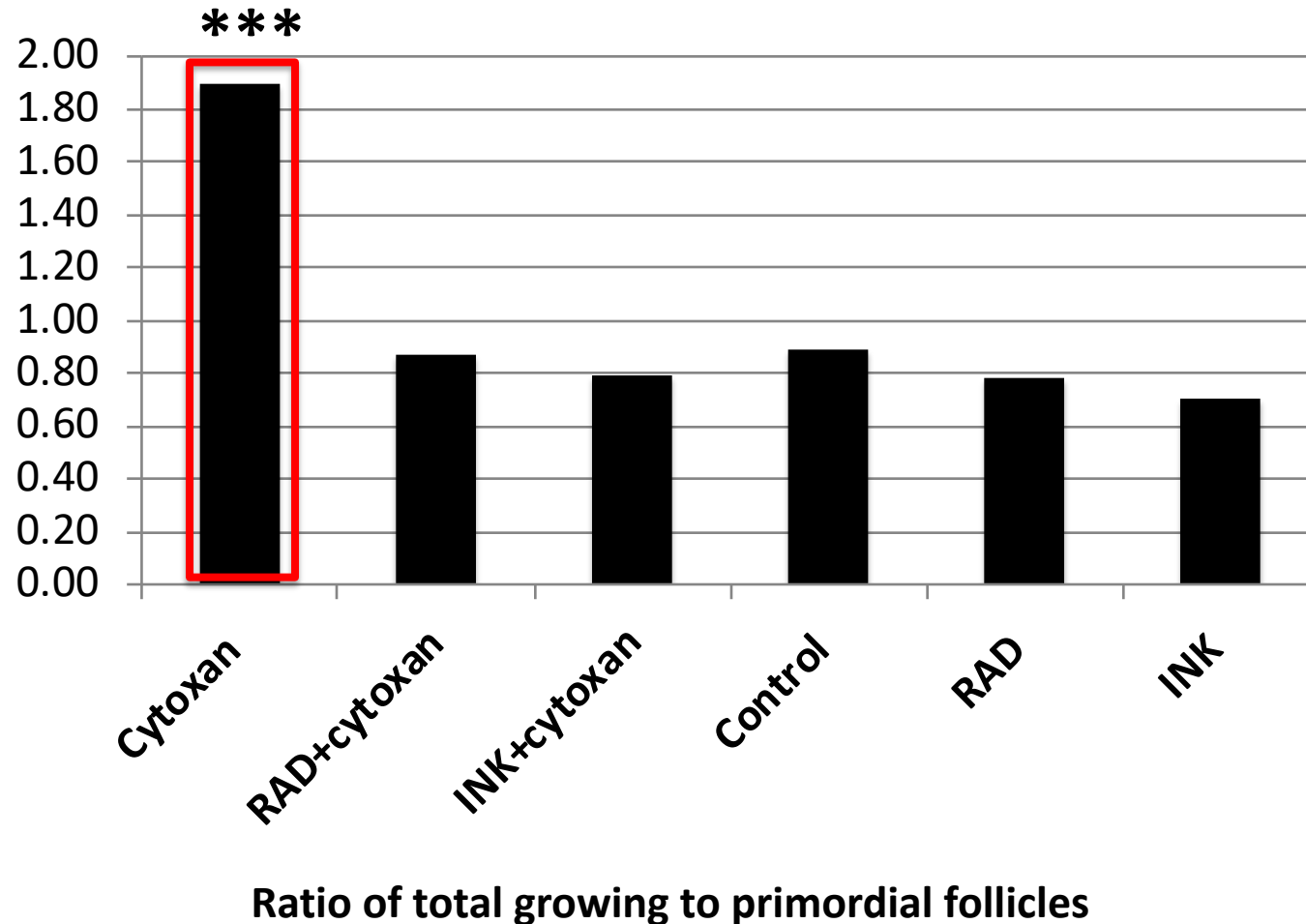
# Phosphorylation of 4EBP-1 and S6 kinase within primordial follicles is decreased after mTOR inhibition



# Two-fold increase in PMFs per surface area when CY-treated mice are co-treated with mTOR inhibitors



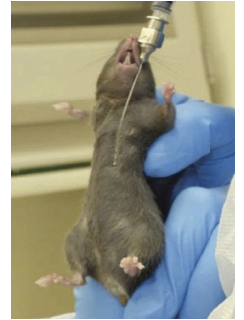
Cytoxan causes follicular burn-out; co-treatment with RAD and INK attenuate this effect





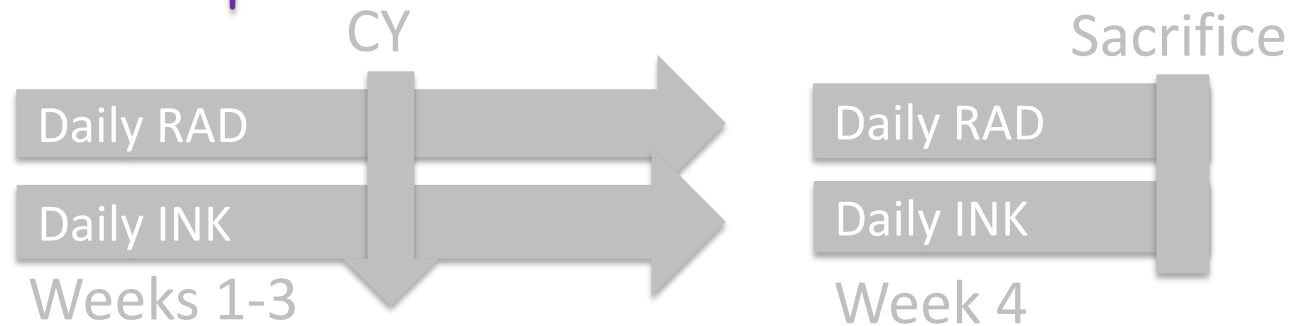
C57BL/6  
8 weeks

Control (PVP) \*  
RAD001\* (mTORC1 inhibitor)  
INK128 \* (mTORC1/2 inhibitor)  
Cyclophosphamide (CY 75mg/kg)  
RAD + CY  
INK + CY

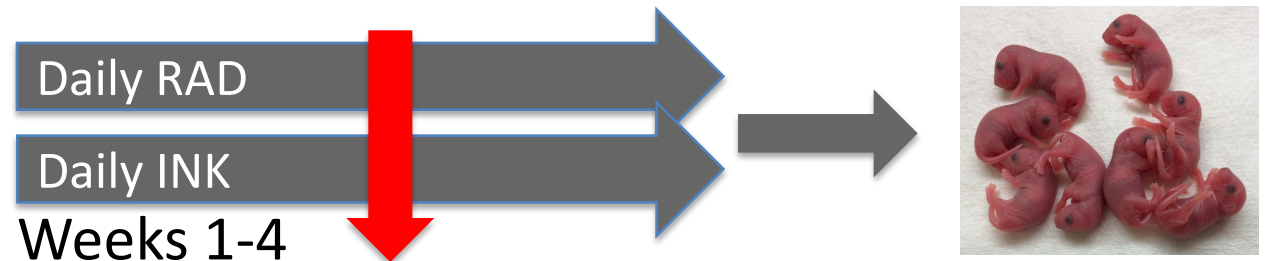


\* Oral gavage

**Aim 1:**  
Ovarian reserve



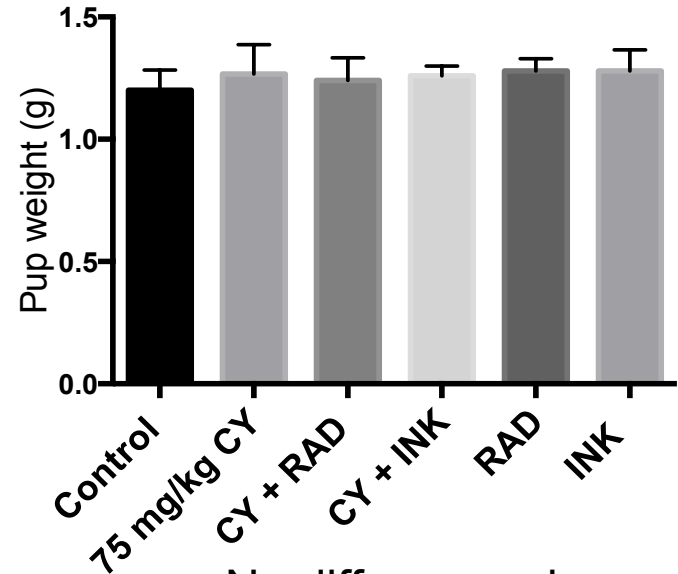
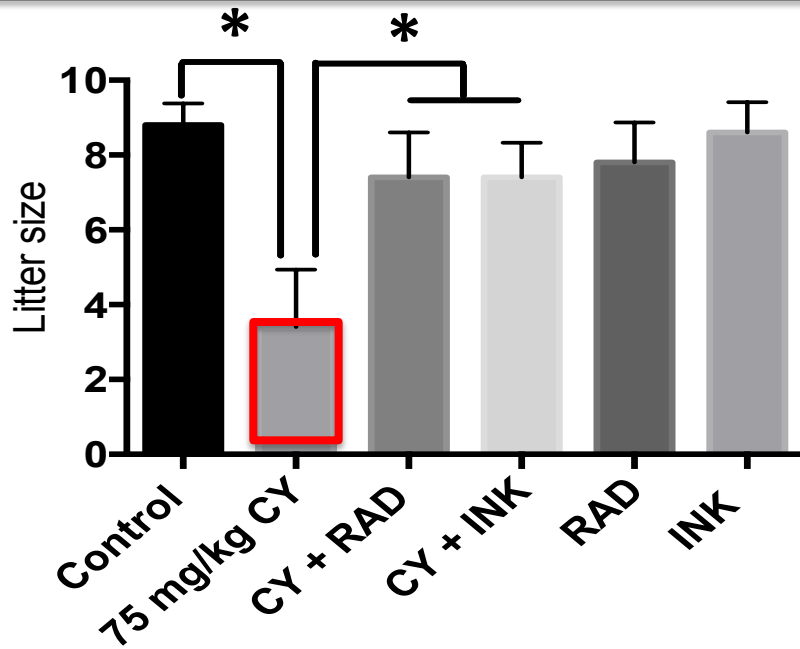
**Aim 2:**  
Fertility





# mTORC1/2 inhibition preserves ovarian function and fertility during genotoxic chemotherapy

Kara N. Goldman<sup>a</sup>, Devon Chenette<sup>b</sup>, Rezina Arju<sup>b</sup>, Francesca E. Duncan<sup>c</sup>, David L. Keefe<sup>a</sup>, Jamie A. Grifo<sup>a</sup>, and Robert J. Schneider<sup>b,d,1</sup>



No differences in pup weight or anomalies



CY

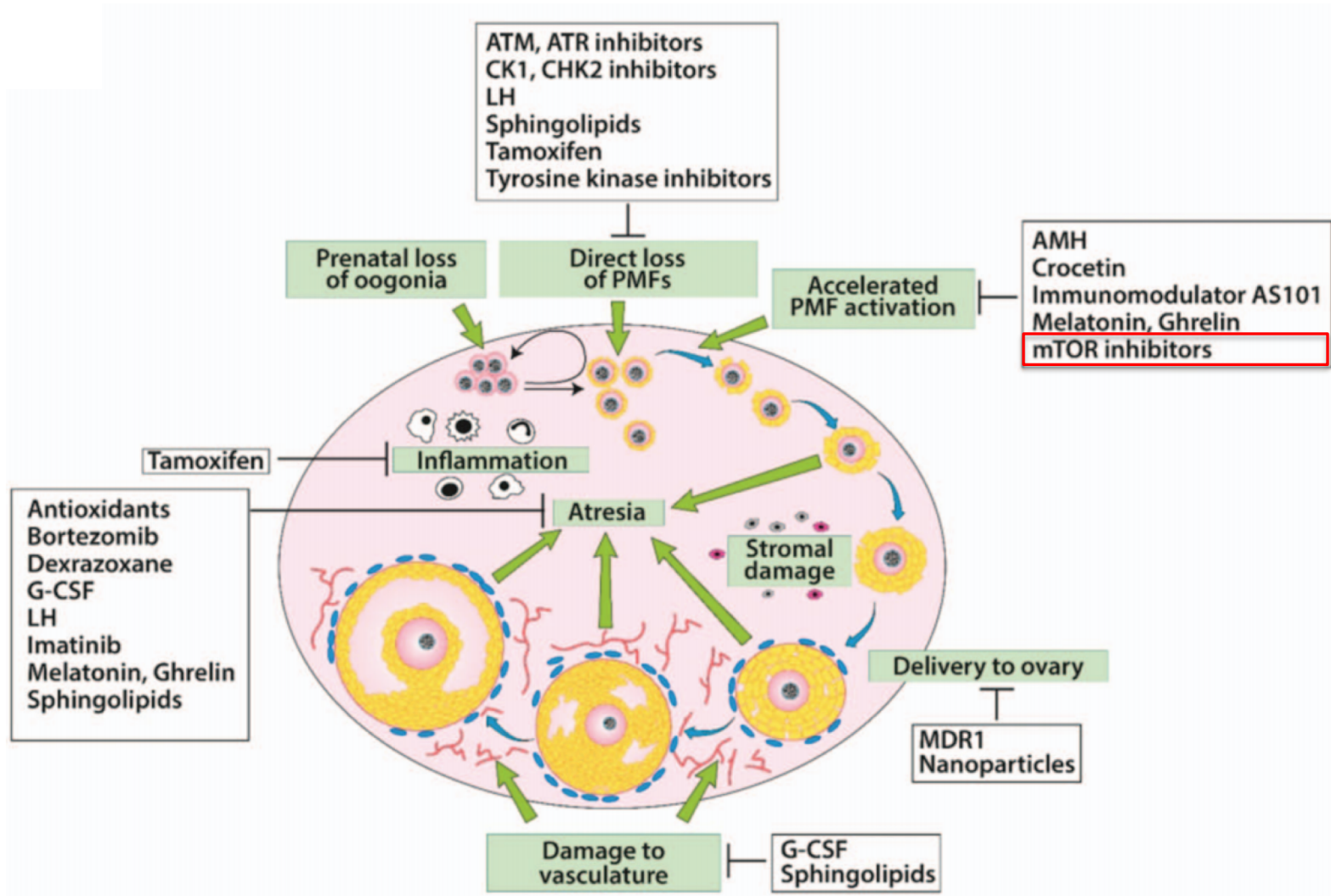


CY + RAD

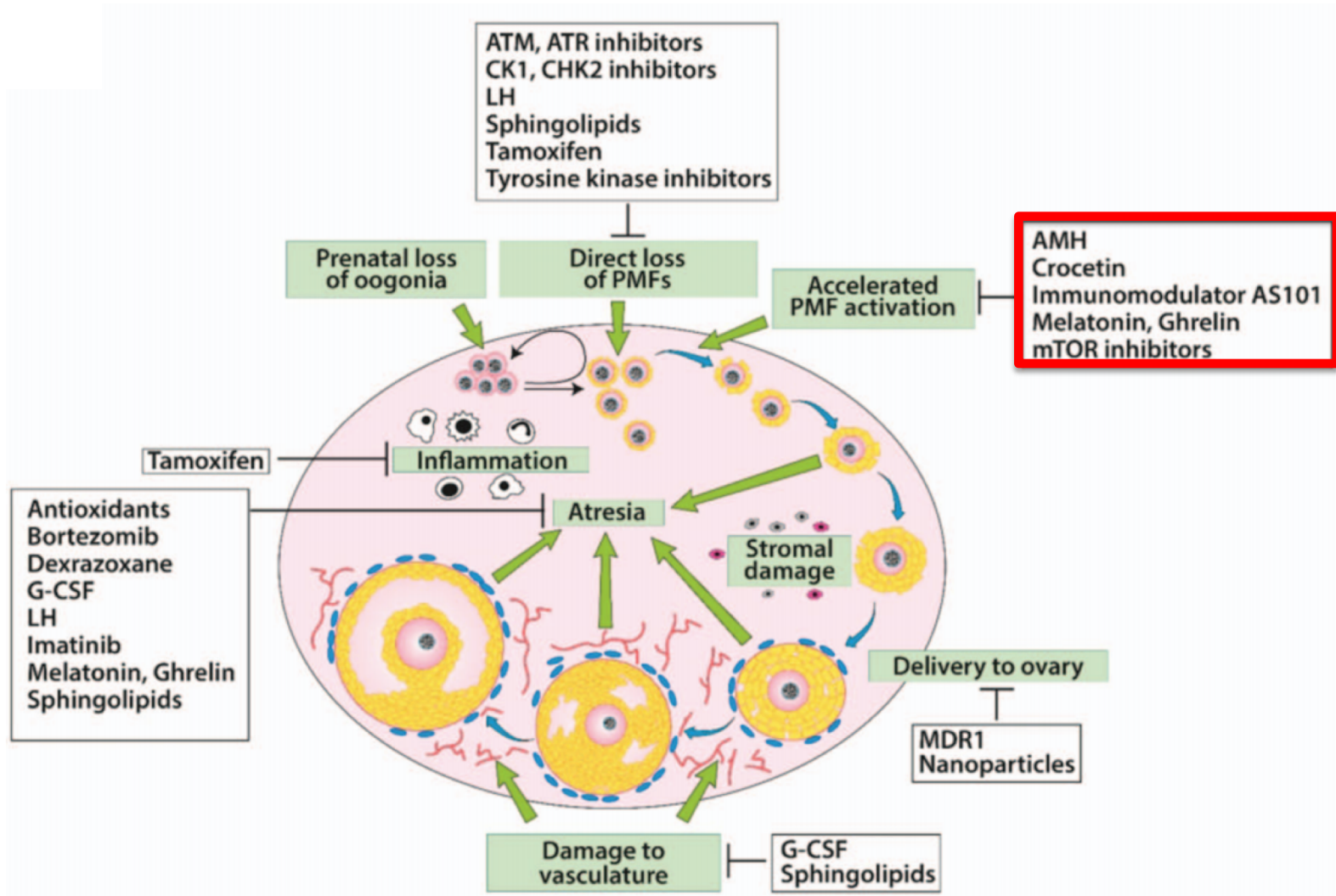


CY + INK

# Growing landscape of pharmacologic fertoprotection



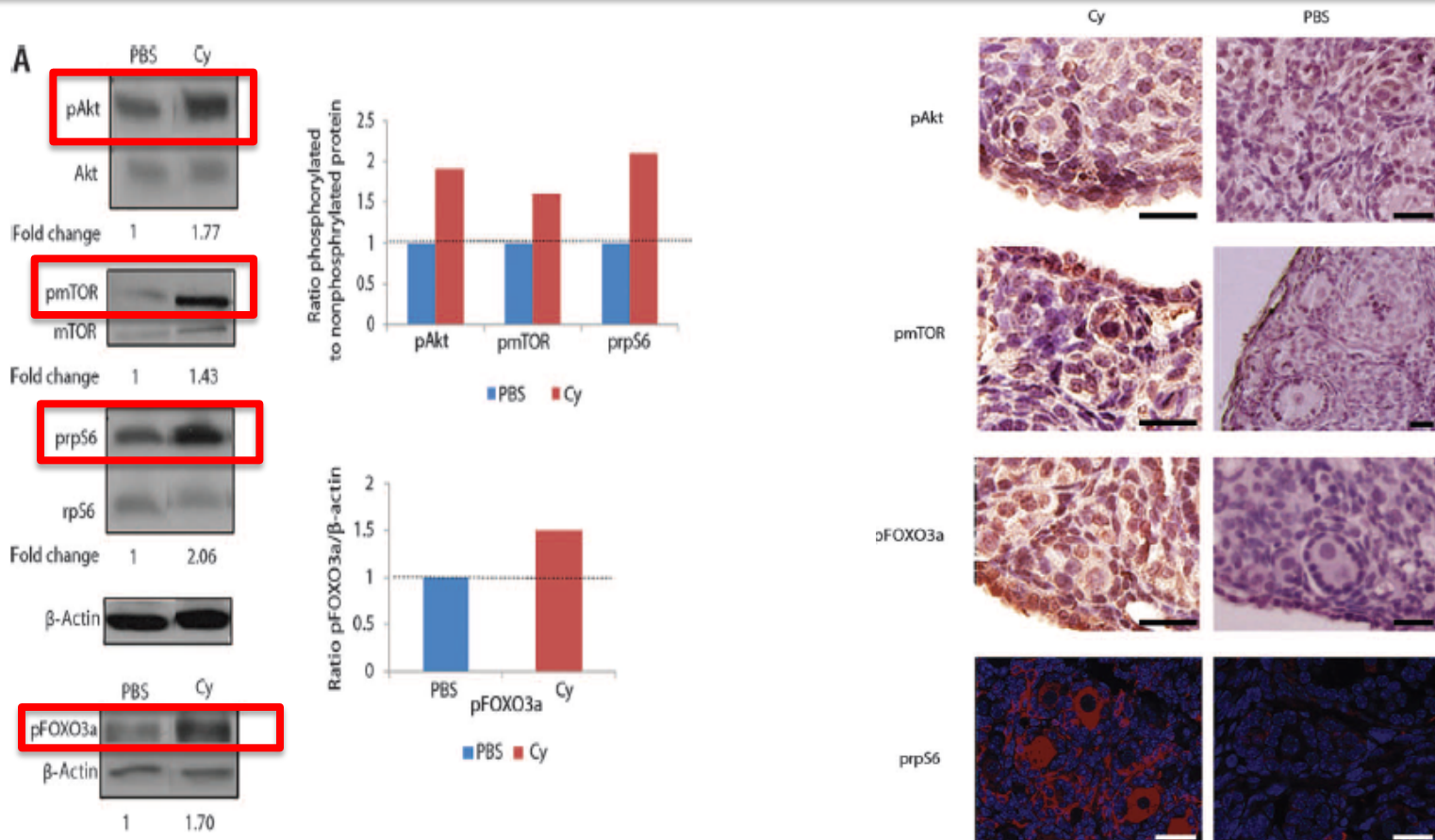
# Growing landscape of pharmacologic fertoprotection



# Cyclophosphamide Triggers Follicle Activation and "Burnout"; AS101 Prevents Follicle Loss and Preserves Fertility

Lital Kalich-Philosoph,<sup>1,2\*</sup> Hadassa Roness,<sup>1\*</sup> Alon Carmely,<sup>1,2</sup> Michal Fishel-Bartal,<sup>1,3</sup> Hagai Ligumsky,<sup>3,4</sup> Shoshana Paglin,<sup>1</sup> Ido Wolf,<sup>3,4</sup> Hannah Kanety,<sup>5</sup> Benjamin Sredni,<sup>2\*</sup> Dror Meiorov<sup>1,3,\*†</sup>

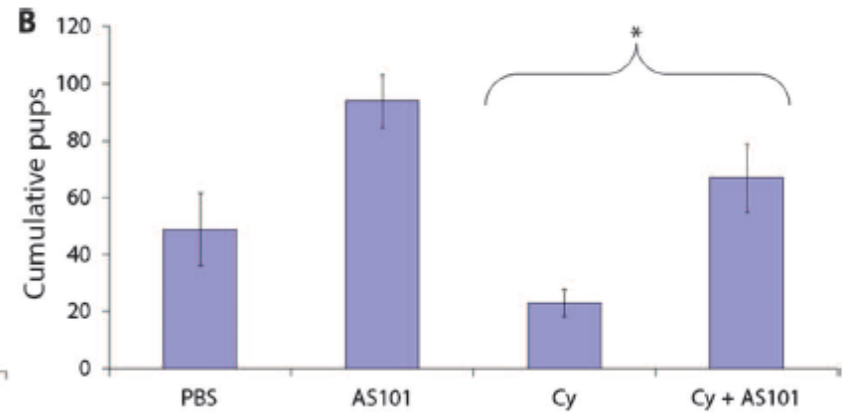
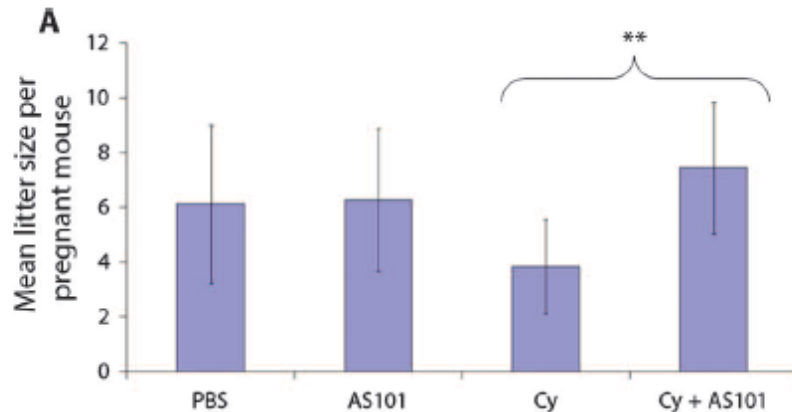
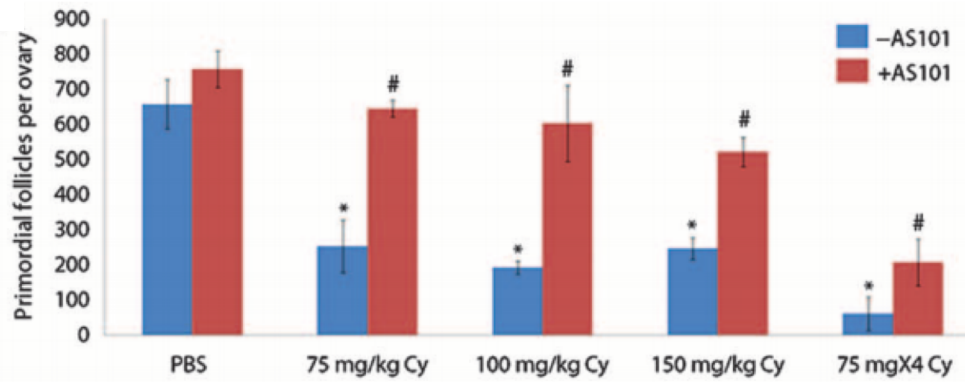
www.ScienceTranslationalMedicine.org 15 May 2013 Vol 5 Issue 185 185ra62



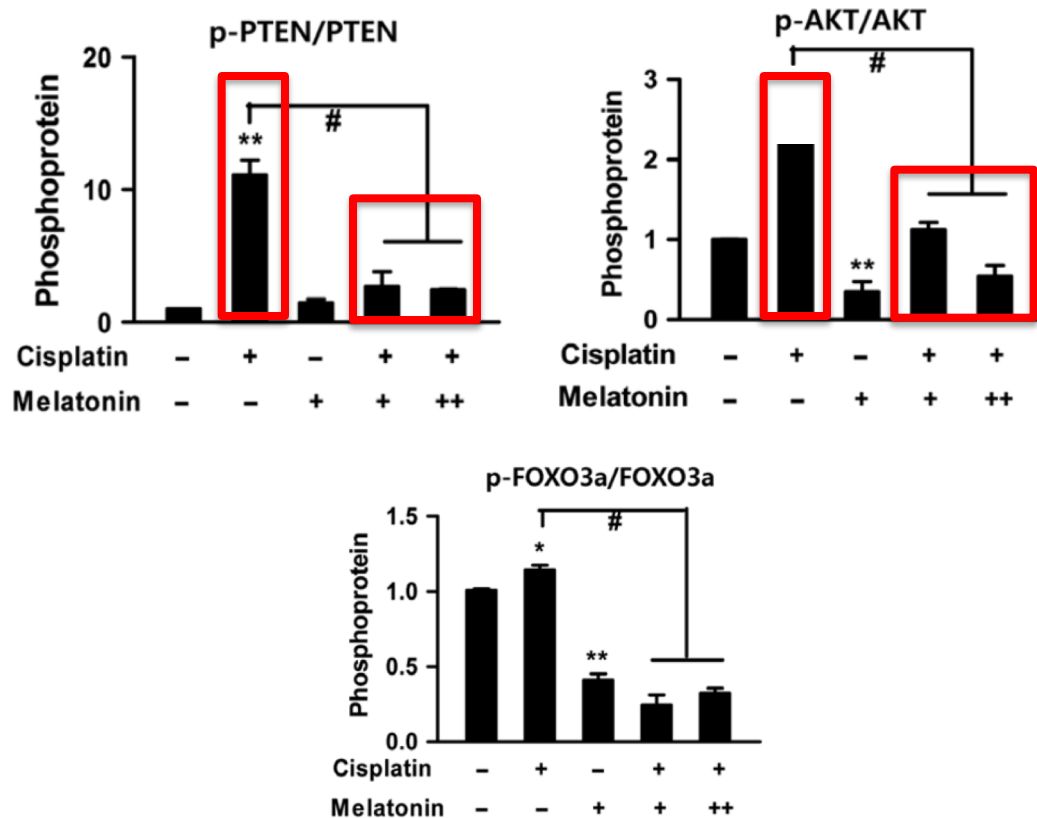
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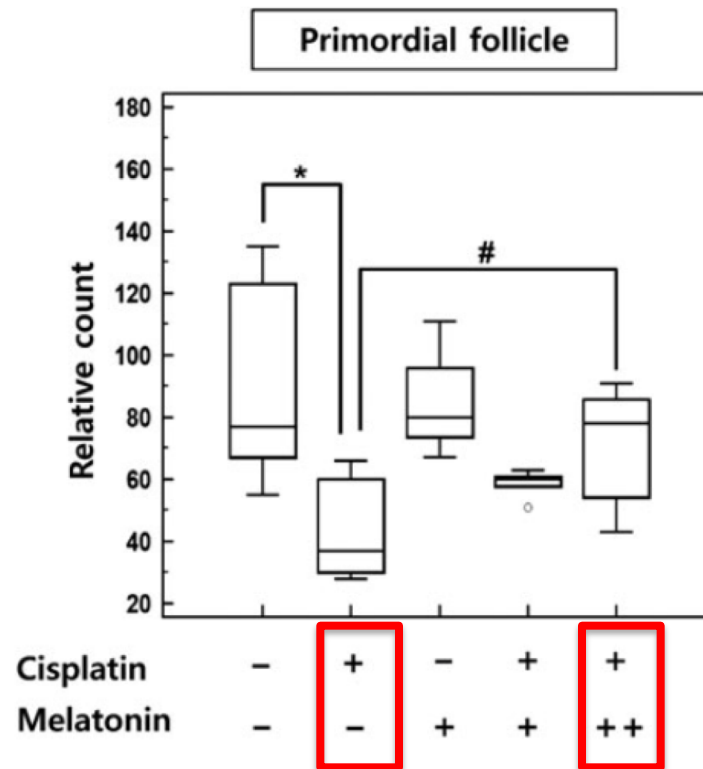
www.ScienceTranslationalMedicine.org 15 May 2013 Vol 5 Issue 185 185ra62



# Melatonin prevents cisplatin-induced primordial follicle loss via suppression of PTEN/AKT/FOXO3a pathway activation in the mouse ovary



# Melatonin prevents cisplatin-induced primordial follicle loss via suppression of PTEN/AKT/FOXO3a pathway activation in the mouse ovary

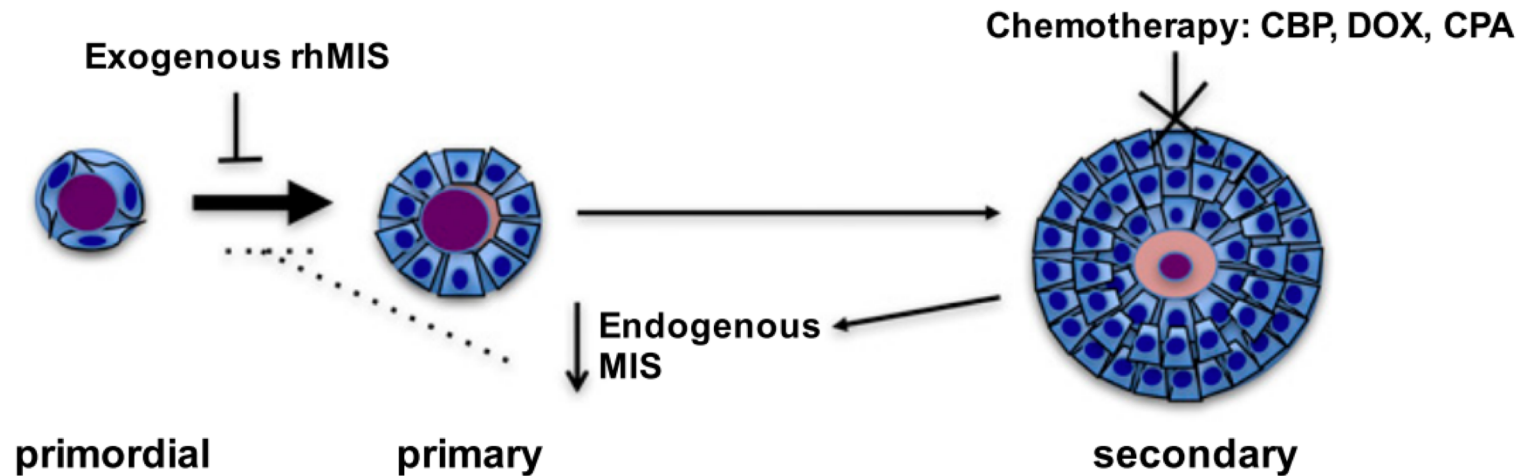


# AMH/MIS as a contraceptive that protects the ovarian reserve during chemotherapy

Motohiro Kano<sup>a,b</sup>, Amanda E. Sosulski<sup>a,b</sup>, LiHua Zhang<sup>a,b</sup>, Hatice D. Saatcioglu<sup>a,b</sup>, Dan Wang<sup>c</sup>, Nicholas Nagykerly<sup>a,b</sup>, Mary E. Sabatini<sup>d</sup>, Guangping Gao<sup>c</sup>, Patricia K. Donahoe<sup>a,b,1</sup>, and David Pépin<sup>a,b,1</sup>

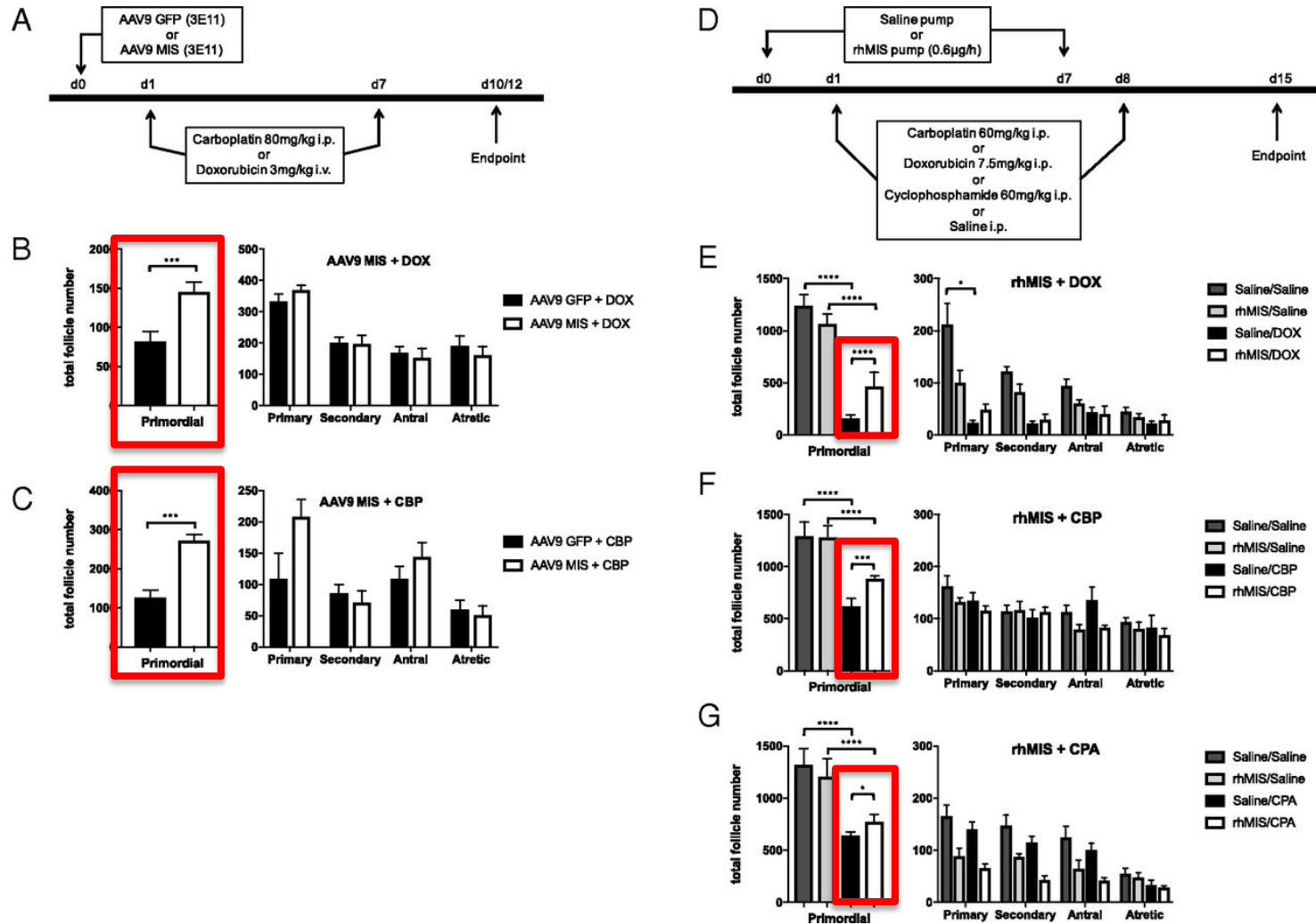
<sup>a</sup>Pediatric Surgical Research Laboratories, Massachusetts General Hospital, Boston, MA 02114; <sup>b</sup>Department of Surgery, Harvard Medical School, Boston, MA 02115; <sup>c</sup>Horae Gene Therapy Center, University of Massachusetts Medical School, Worcester, MA 01655; and <sup>d</sup>Department of Obstetrics and Gynecology, Harvard Medical School and Massachusetts General Hospital, Boston, MA 02114

Contributed by Patricia K. Donahoe, December 29, 2016 (sent for review December 16, 2016; reviewed by Richard N. Freiman, Bruce D. Murphy, and Teresa K. Woodruff)



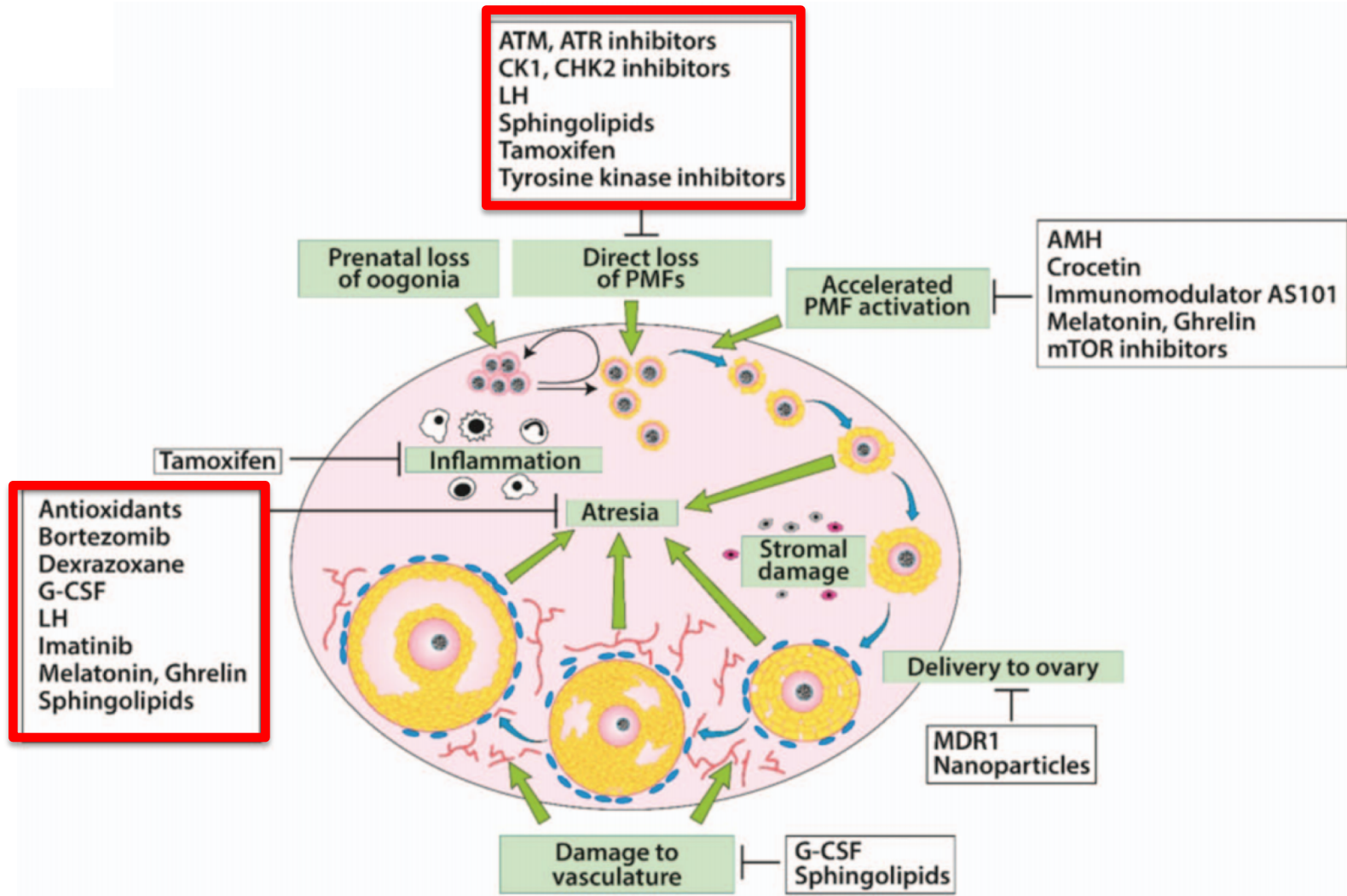


# Treatment with MIS protects the ovarian reserve from the primordial follicle depletion induced by chemotherapy




Motohiro Kano et al. PNAS 2017;114:9:E1688-E1697

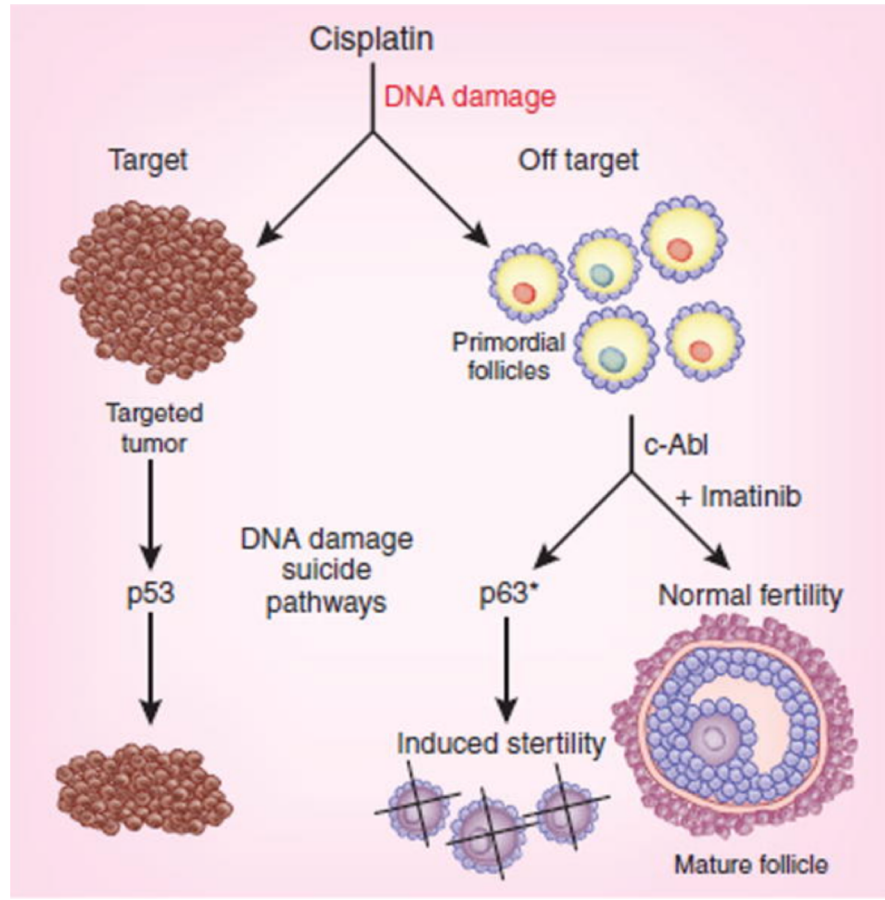
# Growing landscape of pharmacologic fertoprotection



# Inhibition of the c-Abl-TAp63 pathway protects mouse oocytes from chemotherapy-induced death

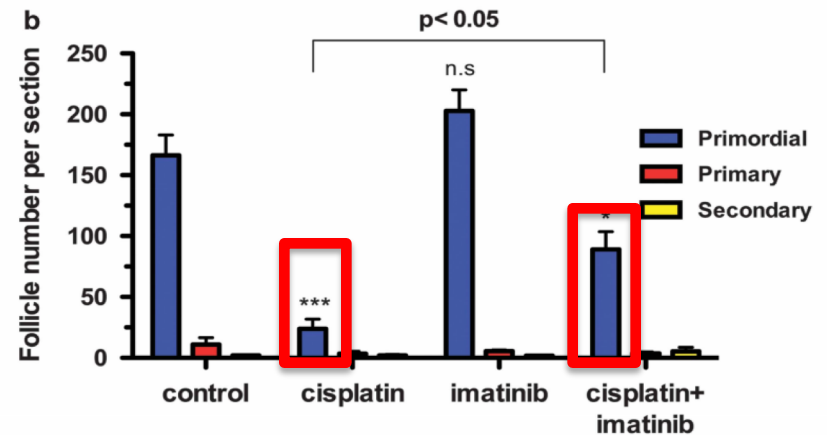
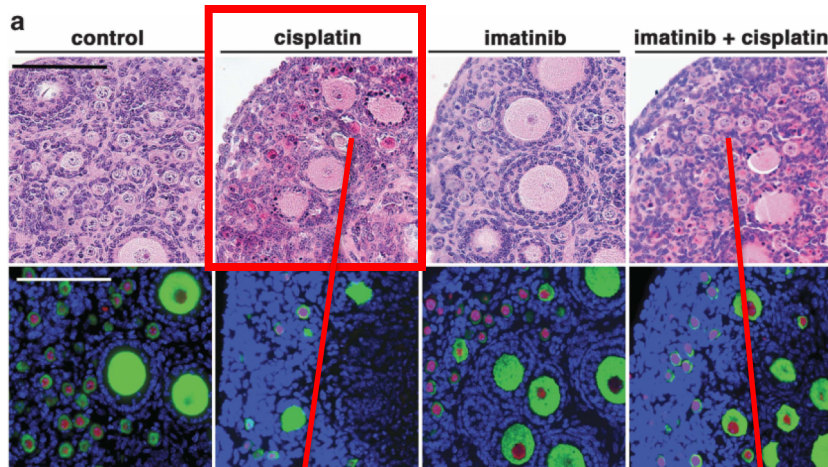
Stefania Gonfloni , Lucia Di Tella, Sara Caldarola, Stefano M Cannata, Francesca G Klinger, Claudia Di Bartolomeo, Maurizio Mattei, Eleonora Candi, Massimo De Felici, Gerry Melino & Gianni Cesareni

*Nature Medicine* **15**, 1179–1185(2009) | [Cite this article](#)



# Rescue of platinum-damaged oocytes from programmed cell death through inactivation of the p53 family signaling network

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Cisplatin-treated ovaries with damaged follicles (pink cytoplasm, condensed nuclei)

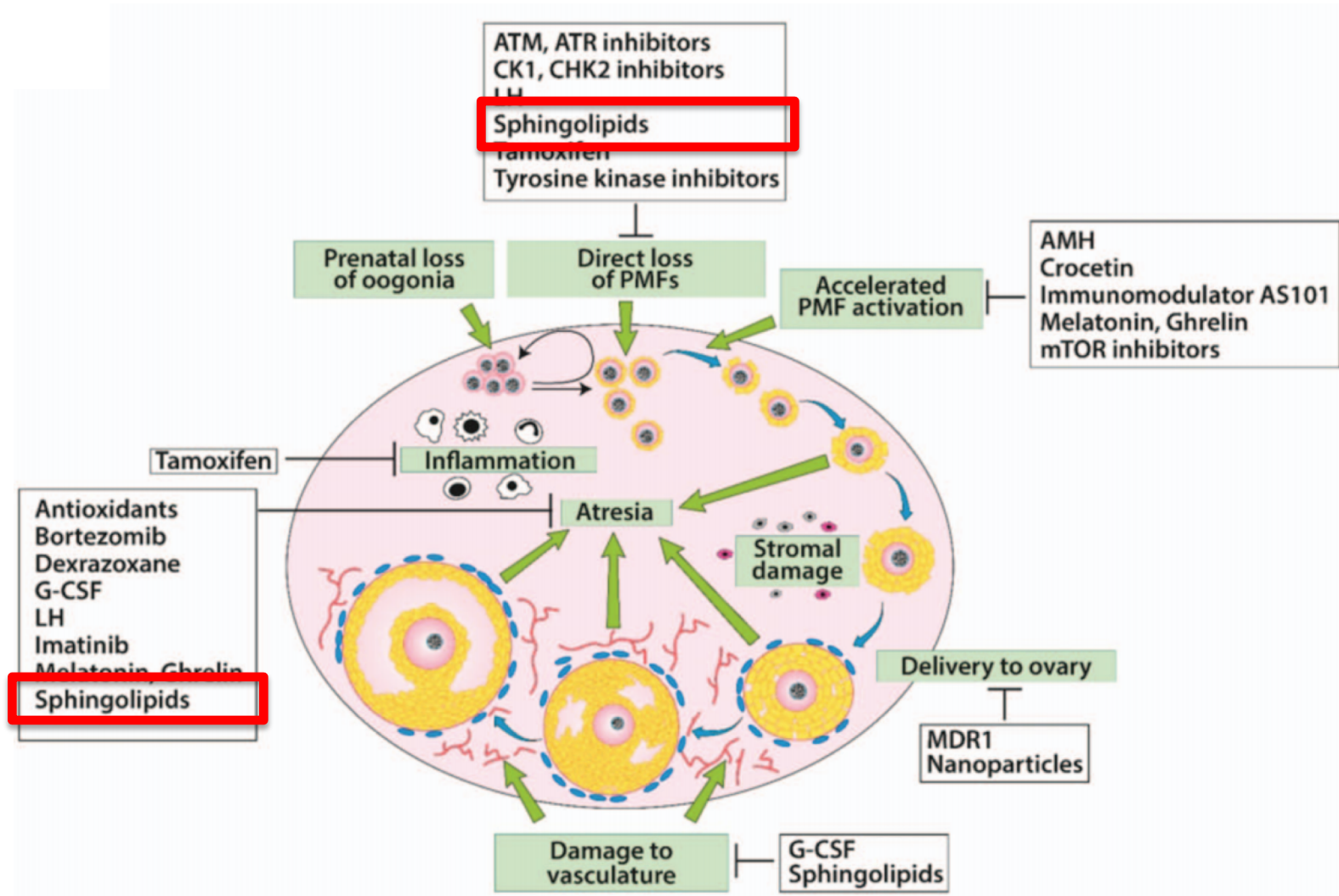
Co-treated mice with healthy appearing PMFs

## **Inhibitors of apoptosis protect the ovarian reserve from cyclophosphamide.**

Luan Y<sup>1</sup>, Edmonds ME<sup>1</sup>, Woodruff TK<sup>1</sup>, Kim SY<sup>1,2</sup>.

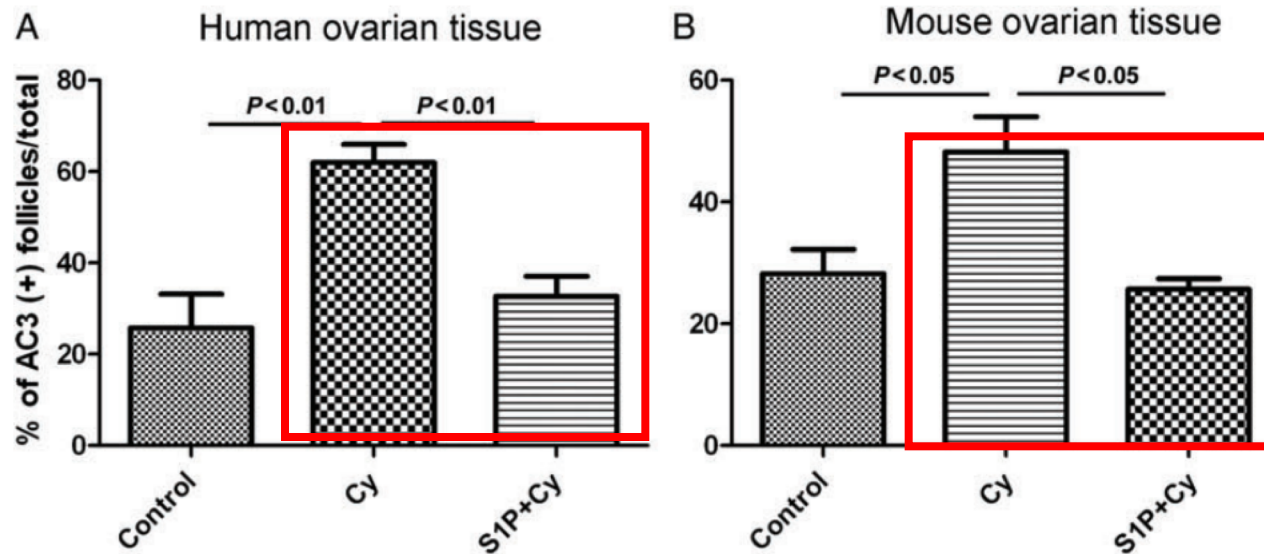
- In vitro evaluation of the effect of CY metabolites in 3 murine strains
  - identified primordial follicle apoptosis
  - Identified phospho-AKT and cleaved PARP within primordial oocytes 3 days after CY injection

# Growing landscape of pharmacologic fertoprotection



# Sphingosine-1-phosphate prevents chemotherapy-induced human primordial follicle death

Fang Li<sup>1,2</sup>, Volkan Turan<sup>1,2</sup>, Sylvie Lierman<sup>3</sup>, Claude Cuvelier<sup>4</sup>,  
Petra De Sutter<sup>3,†</sup>, and Kutluk Oktay<sup>1,2,†\*</sup>

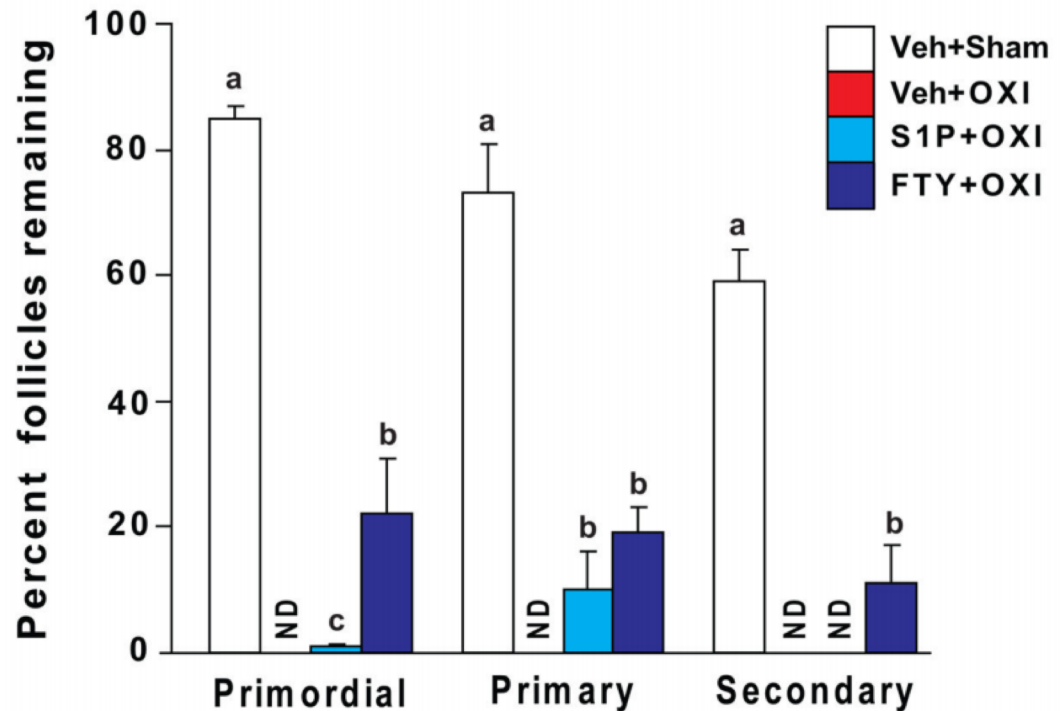


S1P mediated  
CY-induced  
apoptosis

# ***In-vivo* delivery of FTY720 prevents radiation-induced ovarian failure and infertility in adult female non-human primates**

Mary B. Zelinski, Ph.D.<sup>a</sup>, Mark K. Murphy, M.S.<sup>b</sup>, Maralee S. Lawson, B.S.<sup>a</sup>, Andrea Jurisicova, Ph.D.<sup>c</sup>, K. Y. Francis Pau, Ph.D.<sup>a</sup>, Natalia P. Toscano, B.S.<sup>a</sup>, Darla S. Jacob, B.S.<sup>d</sup>, John K. Fanton, D.V.M.<sup>d</sup>, Robert F. Casper, M.D.<sup>c</sup>, Stephen D. Dertinger, Ph.D.<sup>e</sup>, and Jonathan L. Tilly, Ph.D.<sup>f</sup>

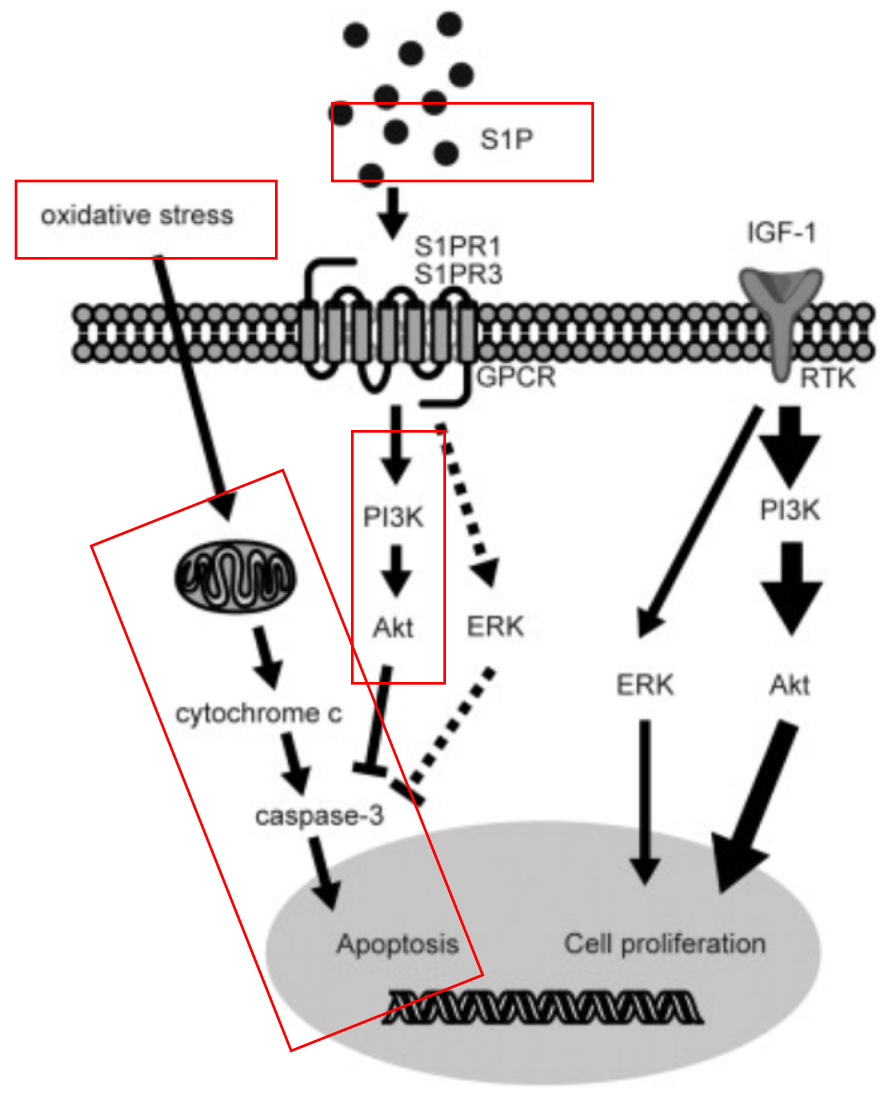
- Intra-bursal S1P and S1P mimetic FTY720 via osmotic mini-pump
  - attenuates radiation-induced oocytes loss in primates





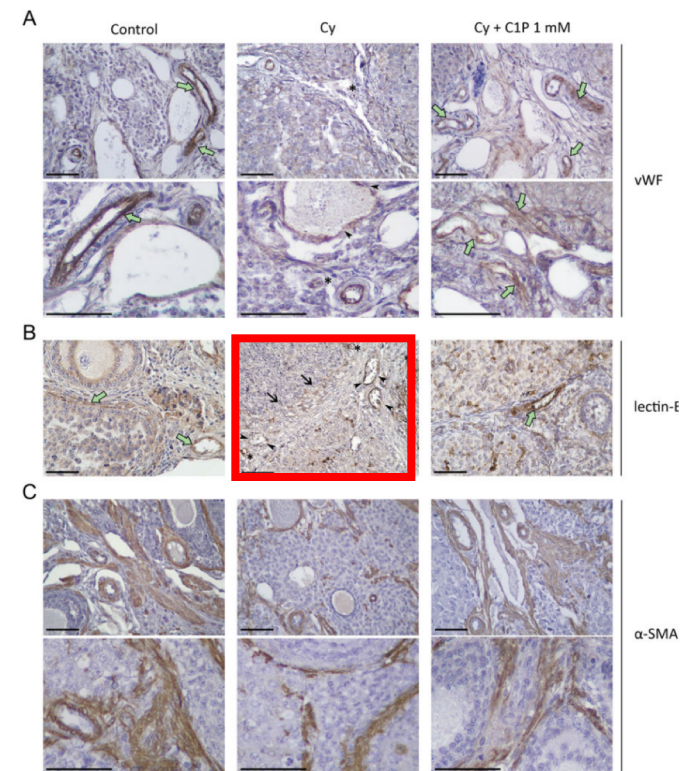
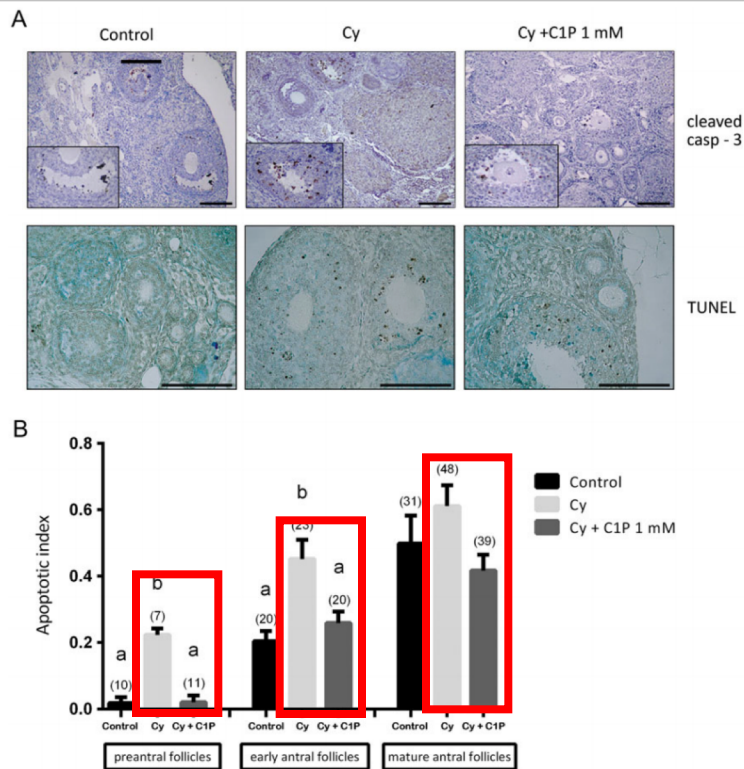
# Oxidative stress, apoptosis, and mTOR

- Oxidative stress induces mitochondrial dysfunction; leads to activation of caspase-3
- S1P may inhibit oxidative stress-induced granulosa cell apoptosis
  - suppressing caspase-3 release via PI3K/AKT signaling pathway



# Ceramide-1-phosphate has protective properties against cyclophosphamide-induced ovarian damage in a mice model of premature ovarian failure

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 Gonzalo Oubiña<sup>1</sup>, Diana Bas<sup>1</sup>, María May<sup>2</sup>, Antonio Gómez Muñoz<sup>3</sup>,  
 Patricia S. Cuasnicú<sup>1</sup>, Débora J. Cohen<sup>1</sup>, Marta Tesone<sup>1</sup>,  
 Dalhia Abramovich<sup>1</sup>, and Fernanda Parborell<sup>1,\*</sup>

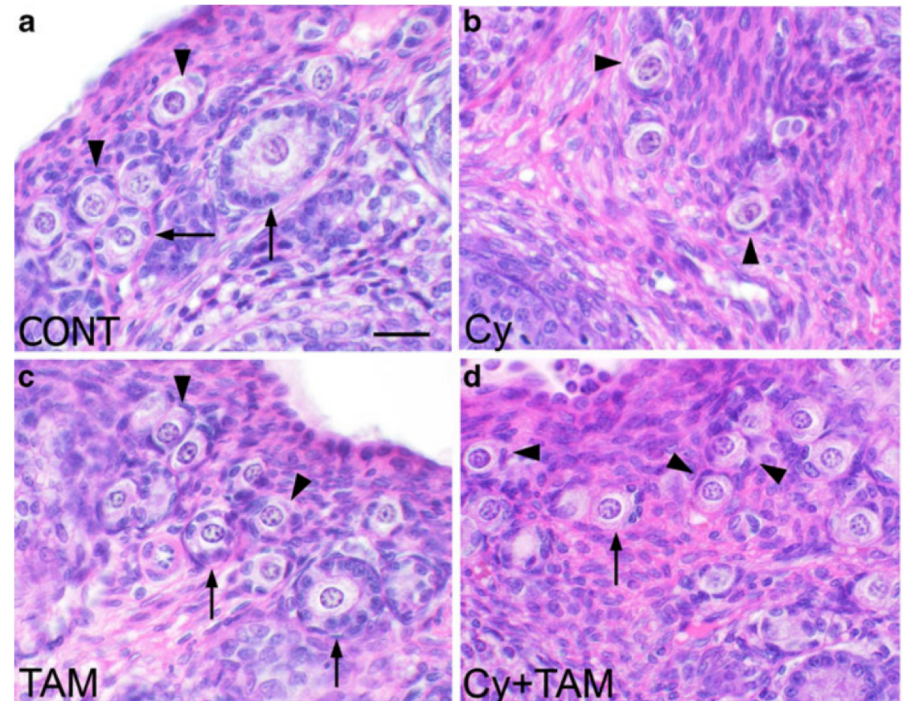
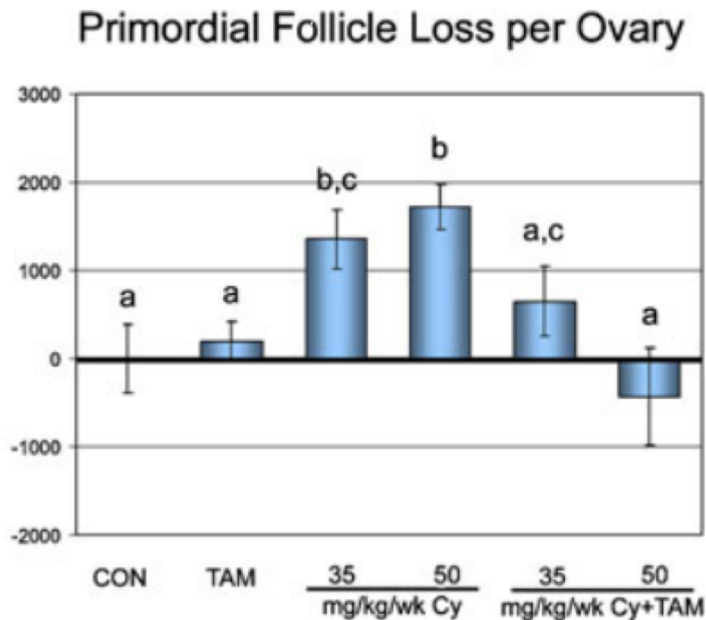


C1P decreased CY-induced apoptosis and reduced CY-induced stromal vascular damage



## Tamoxifen decreases ovarian follicular loss from experimental toxicant DMBA and chemotherapy agents cyclophosphamide and doxorubicin in the rat

Alison Y. Ting · Brian K. Petroff



# Rapidly growing field of fertoprotection

**Table II Agents used to protect ovaries from chemotherapy-induced damage.**

Protectant	Drug	Target action	Species	Reference
AMH/MIS	CPM	Accelerated PMF activation	Mouse	Kano et al. 2016
	DOX Carboplatin			Sonigo et al. 2018
ATM inhibitors: ETP-46464 KUS5399	CIS	Direct loss of PMFs	Mouse	Tuppi et al. 2018
	DOX			Kim et al. 2018
ATR inhibitors: ETP-46464 AZD6738 AS101	CIS	Direct loss of PMFs	Mouse	Kim et al. 2018
	DOX			Luan et al. 2019
	CPM			
Bortezomib Ceramide-1-phosphate	CPM	Accelerated PMF activation	Mouse	Kalich-Philosoph et al. 2013 Di Emidio et al. 2017
	DOX	Atresia	Mouse	Roti Roti et al. 2014
CHK2 inhibitors: BML277 LY2603618 LY2606368	CPM	Direct loss of PMFs	Mouse	Pascuali et al. 2018
	CIS	Direct loss of PMFs	Mouse	Rinaldi et al. 2017
	DOX			Tuppi et al. 2018
CPM	Luan et al. 2019			
CKI inhibitors: MK-8776 CHIR-124	CIS	Direct loss of PMFs	Mouse	Tuppi et al. 2018
	DOX			
PMF670462 PMF4800567 PMF5006739				
Crocetin	CPM	Accelerated PMF activation	Mouse	Di Emidio et al. 2017
Dexrazoxane	DOX	Atresia	Mouse	Kropp et al. 2015
Ghrelin	CIS	Accelerated PMF activation	Mouse	Jang et al. 2017
G-CSF	CIS	Atresia	Mouse	Skaznik-Wikiel et al. 2013
Imatinib	CIS	Vascularisation	Mouse	Akdemir et al. 2014
		Direct loss of PMFs		Kim et al. 2013
Luteinizing Hormone	CIS	Atresia	Mouse	Maiani et al. 2012
				Zamah et al. 2011
				Rinaldi et al. 2017
				Tuppi et al. 2018
				Gonfloni et al. 2009
MDR1	CPM	Delivery to ovary	Mouse	Kim et al. 2018
				Brayboy et al. 2013; 2017 Salih 2011 Wang et al. 2018
Melatonin	CIS	Accelerated PMF activation	Mouse	Jang et al. 2016
Mesna	CIS	Atresia	Rat	Li et al. 2013
Mirtazapine	CIS	Atresia	Rat	Altuner et al. 2013
mTORC inhibitors:	CPM	Accelerated PMF activation	Mouse	Adhikari et al. 2013

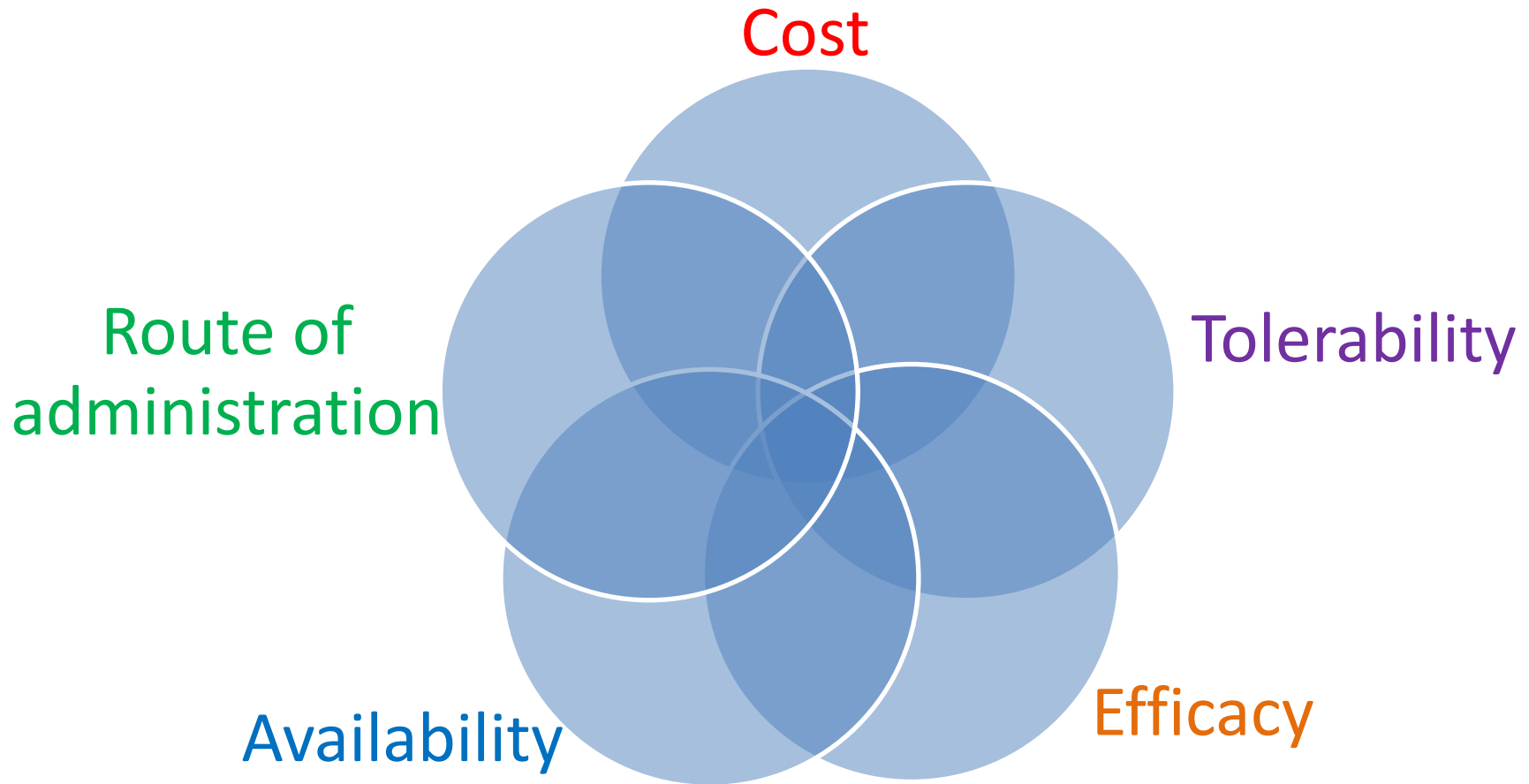
**Table II Continued**

Protectant	Drug	Target action	Species	Reference
Everolimus (RAD001)	CIS			Goldamn et al. 2017
INK128				Zhou et al. 2017
Rapamycin				Tanaka et al. 2018
Resveratrol	CIS	Atresia	Rat	Ozcan et al. 2015
Sphingosine-1-phosphate	CPM	Direct loss of PMFs	Mouse	Morita et al. 2000
			Rat	Li et al. 2017
			Human	Li et al. 2014
				Meng et al. 2014
Sildenafil Citrate	CIS	Atresia	Rat	Taskin et al. 2015
Tamoxifen	CPM	Direct loss of PMFs	Rat	Ting and Petroff 2010
		Inflammation	Human	Piasecka-Srader et al. 2015 Sverrisdottir et al. 2009 Sverrisdottir et al. 2011

# Limitations of existing data

- Pre-clinical
- Highly heterogeneous studies
  - Timing of administration
  - Animal models
  - Chemotherapy regimens

# Qualities important in a fertoprotective agent



*Re-purposed drugs?*

# A win-win for women's reproductive health: A nonsteroidal contraceptive and fertoprotective neoadjuvant

Teresa K. Woodruff<sup>a,1</sup>

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(and a word of cautious optimism):

“In the case of a fertoprotective therapy... we may protect the oocyte from death but damage to the germline may persist, increasing the likelihood of birth defects.”

- Teresa K. Woodruff, PhD

# Thank you

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